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A novel lupane triterpene from seeds of Ziziphus jujuba var. spinosa

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Abstract: Objective To study the chemical constituents from the seeds of Ziziphus jujuba var. spinosa, which is used as a sedative herbal medicine in China. Methods The constituents were separated and purified by silica gel column. Their structures were elucidated based on their physicochemical properties and spectral analysis. Results Six compounds were obtained from 95% EtOH extracts in the seeds of Z. jujuba var. spinosa. They were identified as 2α , 3β -dihydroxy-lup-20(29)-en-28-oic acid methyl ester (1), β -sitosterol (I), betulin (I), betulinic acid (N), hexadexanoic acid 2, 3-dihydroxypropyl ester (V), and daucosterol (V). Conclusion Compound I is a novel lupane triterpene named as alphitolic acid methyl ester, compound V is obtained from the seeds of Z. jujuba var. spinosa for the first time.

Key words: the seeds of Ziziphus jujuba Mill. var. spinosa (Bunge) Hu ex H. F. Chow; lupane triterpene; alphitolic acid methyl ester (2α, 3β-dihydroxy-lup-20(29)-en-28-oic acid methyl ester)

酸枣仁中的一种新羽扇豆烷型三萜

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摘 要:目的 研究酸枣仁 Ziziphus jujuba var. spinosa 的化学成分。方法 利用硅胶柱色谱进行分离纯化,通过理化方法及光谱分析鉴定其结构。结果 从酸枣仁乙醇提取液中得到 6 个化合物,分别鉴定为 2α,3β-dihydroxy-lup-20(29)-en-28-oci acid methyl ester (I)、β-谷甾醇(I)、白桦酯醇(I)、白桦酯酸(N)、1-十六烷酸甘油酯(V)、胡萝卜苷(N)。结论 化合物 I 为新化合物,命名为罗珠子酸甲酯(alphitolic acid methyl ester),化合物 V 为首次从酸枣仁中分离得到。

关键词:酸枣仁;羽扇烷三萜;罗珠子酸甲酯

中图分类号:R284.1 文献标识码:A 文章编号:0253-2670(2006)02-0168-04

The seeds of Ziziphus jujuba Mill. var. spinosa (Bunge) Hu ex. H. F. Chow are used as a sedative medicine in China. There were many published papers which described the isolation methods of new saponins^[1,2], flavonoids^[3], and alkaloids^[4,5]. Here, we present the isolation and structure elucidation of a new lupane triterpene, 2α , 3β -dihydroxy-lup-20(29)-en-28-oic acid methyl ester

(I) named as alphitolic acid methyl ester. Five known compounds β-sitosterol (I), betulin (II), betulinic acid (N), hexadexanoic acid 2, 3-dihydroxypropyl ester (V), daucosterol (V) have also been isolated. Compound V was obtained from Ziziphus jujuba var. spinosa for the first time. The structures of these compounds were elucidated by the means of MS, NMR and physical constant.

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1 Experiment

1.1 Apparatus used

Melting points were determined by an X₄ apparatus. IR spectra were obtained by a Shimadzu Ftir — 8400s spectrometer in KBr-disk. ¹H-NMR, ¹³C-NMR, DEPT, ¹H-¹H COSY, HMQC, and HMBC were recorded with a Brucker AV 400 instrument, with TMS as an internal standard. ESI-MS were recorded in a Quattro Micro™ API instrument. EI-MS was recorded in a JEOL JMS D—300 instrument and HRFTICRMS was measured on a Bruker APEX- I spectrometer. Silica gel (200—300 meshes) was purchased from Qingdao Marine Chemical Factory. TLC were performed on HPTLC plates (Alltech).

1.2 Plant material

The seeds of Z. jujuba var. spinosa (11.0 kg) were collected, from Jixian, Tianjin of China in November 2002 and were identified by Professor Qin Minjian, Department of Natural Resource, China Pharmaceutical University. A retained specimen (Specimen No. z-007-2) has been deposited in Tianjin Zhongxin Pharmaceuticals, Tianjin, China.

1.3 Extraction and isolation

The seeds of Z. jujuba var. spinosa were extracted with 95% EtOH (3×20 L). After removal of the solvent under reduced pressure, the extract was dissolved in water and extracted with petroleum, EtOAc, and n-BuOH, respectively. The EtOAc extract (60 g) was fractioned by silica gel column (200 – 300 meshes, 500 g) with gradient elution of CHCl₃-CH₃OH and purified by silica gel column (200 – 300 meshes) eluted with petroleum-EtOAc to yield I (12 mg), I (15 mg), I (5 mg), N (1.89 g), V (9 mg) and V (10 mg).

2 Results and discussion

Compound I, mp 248 – 251 °C, colorless needles in MeOH, ESI-MS m/z 487.4 [M+H]⁺ and a sodium molecular ion [M + Na] at m/z 509.716 3 (calcd. 509.716 1) in the HRFTI-CRMS, corresponding with a molecular formula of $C_{31}H_{50}O_4$. IR ν_{max}^{KBr} cm⁻¹: 3 387, 3 070, 2 947, 1 724, 883. The molecular formula $C_{31}H_{50}O_4$ im-

plies seven degrees of unsaturation. It gave positive response to Libermann-Burchard reaction, that suggested that it should be triterpene. The ¹H-NMR spectrum showed five angular methyl groups [δ 0. 98 (s, C₂₃-H₃), 0. 94 (s, C₂₇-H₃), 0. 89 (s, C₂₆-H₃), 0. 87 (s, C₂₅-H₃), 0. 78 (s, C₂₄-H₃)], and one allyl group [δ 1. 66 (s, C₂₉-H₃); 4. 71, 4. 58 (each br s, C₃₀-H₂)]. The ¹³C-NMR and DEPT showed 31 carbon signals which included one carbonyl carbon [δ 176. 64 (s) in ¹³C-NMR] and one methoxy group [δ 51. 25 (q) in ¹³C-NMR and δ 3. 64 (s) in ¹H-NMR] ¹H-NMR and ¹³C-NMR were submitted in Table 1.

Table 1 ¹H-NMR and ¹³C-NMR Data of betulinic acid and compound I in CDCl₃

No.	Betulinic acid (100 MHz) ^[6] C	Compound 1	
		С	Н
1	39. 0	46.77 (t)	2.02 (dd, J=12.0,4.8)
			0.82 (t, $J=12.0$)
2	27.6	69.25 (d)	3.64 (m)
3	78. 2	83.91 (d)	2.94 (d, $J=12.0$)
4	39.0	39.18 (s)	_
5	55. 5	55.47 (d)	0.77 (m)
6	18. 4	18.28 (t)	1.48 (m),1.34 (m)
7	34.5	34.21 (t)	1.34 (m)
8	40.8	40.75 (s)	
9	50.7	50.49 (d)	1.30 (m)
10	37.3	38.58 (s)	_
11	21.0	21.00 (t)	1.43 (m)
12	25. 6	25.40 (t)	1.69 (m),1.00 (m)
13	38. 2	38.20 (d)	2.15 (m)
14	42.5	42.45 (s)	_
15	30.8	30.61 (t)	1.87 (m),1.33 (m)
16	32.6	32.16 (t)	2.20 (m),1.37 (m)
17	56. 3	56.55 (s)	_
18	47.1	46.97 (d)	2. 96 (t, $J = 11.2$)

Because of the long-range correlations showed between the carbonyl carbon and the methoxy group in HMBC spectrum (Fig. 1), the methoxy group was connected to the carbonyl carbon. The 13 C-NMR spectra of I was in a good agreement with those of betulinic acid [6] except the C-28 and the carbons in the ring A (Table 1). The IR absorption of I at 3 387 cm⁻¹ showed the presence of hydroxyl groups. Comparing with the molecular formula and degrees of unsaturation, we know that there are two hydroxyl groups in it [δ 69. 25 (d) and 83. 91 (d) in 13 C-NMR]. With the HMBC spectrum (Fig. 1), we conclused that the two hy-

droxyl groups connected to the C-2 and C-3, which was also been proved by $^1\text{H}_-^1\text{H}$ COSY (Fig. 2). From the above, the structure of I was lup-20 (29)-en-28-oic acid, 2, 3-dihydroxy-, methyl ester. In ^1H -NMR spectrum, δ 0.82 (1H, t, J= 12.0 Hz, H-1a) and δ 2.02 (1H, dd, J= 12.0, 4.8 Hz, H-1b) showed the C₂-OH was α configuration, and δ 2.94 (1H, d, J= 12.0 Hz, H-3) showed the C₃-OH was β configuration (Fig. 3). From the above evidence, I was elucidated as 2α , 3β -dihydroxy-lup-20 (29)-en-28-oic acid methyl ester (Fig. 4).

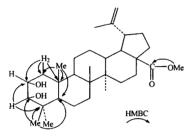
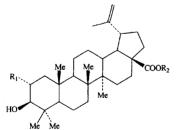


Fig. 1 HMBC Correlations in compound I

SY Correlations in compound I

Fig. 3 Structure of ring A in compound I



Compound I: $R_1=OH$, $R_2=CH_3$ Betulinic acid: $R_1=R_2=H$

Fig. 4 Sturctures of compound I and betulinic acid

Compound II, mp 262-265 °C, colorless-needles in MeOH, it gave positive response to Libermann-Burchard reaction. ESI-MS m/z 442 [M+H]⁺. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹:3 370, 3 090, 1 650,885.

¹H-NMR (CDCl₃) δ 0. 74, 0. 81, 0. 95, 0. 96, 1. 00 (3H×5, s, 23, 24, 25, 26, 27-Me), 1. 66 (3H, s, 30-Me), 3. 31; 3. 77 (2H, each d, J=10. 8 Hz,

H-28), 4. 56; 4. 66 (2H,s,H-29), 3. 16 (1H, dd, J=11.2, 6. 4 Hz, H-3). ¹³C-NMR (CDCl₃) δ 38. 69 (C-1), 27. 36 (C-2), 78. 90 (C-3), 38. 84 (C-4), 55. 28 (C-5), 18. 28 (C-6), 34. 23 (C-7), 40. 91 (C-8), 50. 40 (C-9), 37. 15 (C-10), 20. 82 (C-11), 25. 21 (C-12), 37. 30 (C-13), 42. 71 (C-14), 27. 04 (C-15), 29. 17 (C-16), 47. 77 (C-17), 47. 77 (C-18), 48. 76 (C-19), 150. 45 (C-20), 29. 75 (C-21), 33. 96 (C-22), 27. 97 (C-23), 15. 33 (C-24), 16. 08 (C-25), 15. 96 (C-26), 14. 74 (C-27), 60. 49 (C-28), 19. 06 (C-29), 109. 66 (C-30). Comparing to the reference [6], III was identifed as betulin.

Compound N, mp 287 - 289 °C, colorless needles in MeOH, it gave positive response to Libermann-Burchard reaction. ESI-MS m/z 455. 2 $[M-H]^+$. IR ν_{max}^{KBr} cm⁻¹: 3 450, 1 686, 1 650, 875. ¹H-NMR (CDCl₃) δ 0. 74, 0. 81, 0. 93, 0. 95. $0.96 (3H \times 5, s, 23, 24, 25, 26, 27-Me), 1.68$ (3H,s,30-Me), 4. 60; 4. 72 (2H, each d, J=12.0Hz, H-29), 3.16 (1H, dd, J=11.2, 6.4 Hz, H-3). ¹³C-NMR (CDCl₃) δ 38.71 (C-1), 27.39 (C-2), 79.01 (C-3), 38.85 (C-4), 55.35 (C-5), 18.28 (C-6), 34.32 (C-7), 40.69 (C-8), 50.52 (C-9), 37. 21(C-10), 20. 85(C-11), 25. 50(C-12), 38.40 (C-13), 42.43 (C-14), 30.56 (C-15), 32.16 (C-16), 56.31 (C-17), 46.89 (C-18), 49.28 (C-19), 150.40 (C-20), 29.70 (C-21), 37.02 (C-22), 27.98 (C-23), 15.32 (C-24), 16.10 (C-25), 16.02 (C-26), 14.68 (C-27), 180.70 (C-28), 19.36 (C-29), 109.66 (C-30). Comparing to the reference [6], N was identified as betulinic acid.

Compound V, mp 68-72 C, white powder, EI-MS m/z 299 [M - CH₂OH]⁺, 239 [M - C₃H₇O₃]⁺, 134 [M - C₁₄H₂₈]⁺, 74 [M - OOCC₁₅ H₃₁]⁺, 57 [C₄H₉]⁺, 43 [C₃H₇]⁺. ¹H-NMR (CD-Cl₃) δ 4.19 (1H, dd, J=11.6, 4.8 Hz), 4.13 (1H, dd, J=11.6, 6.0 Hz), 3.91 (m), 3.68 (1H, dd, J=11.2, 4.0 Hz), 3.58 (1H, dd, J=11.2, 6.0 Hz), 2.33 (2H, t, J=7.6 Hz), 1.61 (2H, m, J=7.6 Hz), 1.4-1.2 (24H, m), 0.86 (3H, t, J=6.8 Hz). ¹³C-NMR (CDCl₃) δ 174.32

(s), 70.26 (d), 65.16 (t), 63.31 (t), 34.14 (t), 31.90(t), 30-29(t), 14.08(q). Comparing to the reference^[7], V was identified as hexadexanoic acid 2, 3-dihydroxypropyl ester.

Compound I, mp 139-140 °C, was isolated as colorless needle, and compound VI, mp 292-295 °C, was isolated as colorless powder. They were identified as β-sitosterol and daucosterol, respectively after the determination of the mixed melting point and Rf vs authentic samples.

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络石藤中的三萜类化合物

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摘 要:目的 对络石藤中的三萜类化合物进行分离鉴定。方法 采用硅胶、反相硅胶、Sephadex LH-20 等柱色谱 方法进行分离, NMR 等波谱学方法进行结构鉴定。结果 从络石藤中分离鉴定 8 个三萜类化合物: 络石苷 F(trachelosperoside F, I)、络石苷 B-1(trachelosperoside B-1, I)、络石苷 D-1(trachelosperoside D-1, I)、络石苷 E-1 (trachelosperoside E-1, N), 3β-O-D-glucopyranoside quinovic acid (V), 3β-O-β-D-glucopyranoisde 'quinovic acid 27-O-β-D-glucopyranosyl ester (VI), 3β-O-β-D-glucopyranoside cincholic acid 27-O-β-D-glucopyranosyl ester (VI), 络石苷元 B(trachelosperogenin B, W)。结论 I 为新化合物,其余均为首次从该植物中分离得到。

关键词:络石藤;三萜;trachelosperoside F

中图分类号:R284.1

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Triterpenoids from canes with leaves of Trachelospermum jasminoides

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Abstract: Objective To study the triterpenoids from the canes with leaves of Trachelospermum jasminoides. Methods The compounds were separated and purified by column chromatography with silica gel, RP-C₁₈, Sephadex LH-20 and identified by IR, MS, NMR, and 2D-NMR. Results Eight triterpenoids were identified as: trachelosperoside F (I), trachelosperoside B-1 (I), trachelosperoside D-1 (II), trachelosperoside E-1 (N), 3β-O-D-glucopyranoside quinovic acid (V), 3β-O-β-D-glucopyranoisde

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