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A new triterpenoid saponin from Lysimachia candida

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Abstract Object To investigate the chemical constituents from the whole plant of Lysimachia candida. Ldl. Methods The constituents were isolated and purified on silica gel column chromatography. Their structures were elucidated by chemical and spectroscopic evidence. Results A triterpenoid saponin, named candidoside A (I), was isolated from the extract of n-BuOH. Its structure was shown to be β , 16α -dihydroxy-olean-12-en-28-al-3- $O\beta$ -D-glucopyranosyl-23- $O\alpha$ -D-ribofuranoside. Conclusion Candidoside A was a new triterpenoid saponin.

Key words Lysimachia candida Ldl.; triterpenoid saponin; candidoside A

从单条草中分得一个新的三萜皂苷

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摘 要:目的 对报春花科珍珠菜属药用植物单条草 $Lysimachia\ candida$ 的化学成分进行研究 方法 采用硅胶柱层析进行分离和纯化,通过波谱和化学方法进行结构鉴定。结果 从正丁醇萃取部分分离出 1个三萜皂苷类化合物,结构鉴定为: $33,16\alpha$ —二羟基齐墩果 -12%—-28醛—-3-O3—-D4、喃葡萄糖基—-23-O4、-D4、喃核糖苷,命名为单条草苷甲(I)、结论 单条草苷甲是新结构的三萜皂苷。

关键词: 单条草;三萜皂苷;单条草苷甲

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The plant of *Lysimachia candida* Ldl. (Primulaceae), a Chinese folk medicine, has long been used to treat detoxication and promote blood circulation. We reported one new triterpenoid saponin and 12 known compounds from the methanolic extracts^[1]. This paper deals with the separation and structure elucidation of another new triterpenoid saponin, named as candidoside A (I).

1 Results and Discussion

Candidoside A (I) was obtained as amorphous powder. Its formula was determined as C41 H66O13 by HR-FAB-MS at m/z 767.4557 ([M+ H], calcd. 767.4582). The IR spectrum exhibited absorptions at 3 432 (OH) and 1 710 cm $^{-1}$ (C= O). Acid hydrolysis of I yielded glucose and ribose on TLC. In the positive FAB-MS

ment ion peaks at m/z 605 [M-gle+ H] and 455 [M-gle- rib- H-O+ H] were observed. The EI-MS of acetylated I—showed fragment ion peaks at m/z 259 [rib (O Ac)_3] and 331 [gle (O Ac)_4], suggesting that glucose and ribose moieties should be terminal monosaccharide. In the HNMR and CNMR spectra, the presence of seven quaternary carbon atoms and the chemical shifts of C-12 at δ 123. 6 and C-13 at δ 143. 6 were characteristics of a Δ^{12} -oleanene skeleton. Acid hydrolysis of I—gave aglycone I—a, whose 13 CNMR spectrum disclosed the presence of three hydroxylated carbons (δ 78. 1, 73. 0, 73. 5). The signal of C-3 (δ 78. 1) suggested an equatorial position for the 3-OH.

Comparing the $^{13}\,\text{CN\,M\,R}$ signals of I –a with

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those of oleanolic acid^[2], C-5 of I a was shifted upfield to δ 49. 8. According to the rule of steric effect, a hydroxyl must be linked to C-23 (δ 73. 5). The signal of δ 205. 7 (d) and 73. 1 (d) indicated an aldehyde and a tertiary hydroxyl group. Except for C-17, C-22 and C-28, the ¹³CNMR signals of I a were almost identical with those of 33, 16a, 23, 28-tetrahydroxy-olean-12-ene^[3], which revealed that \alpha -OH and -CHO were linked to C-16 and C-17, respectively. This conclusion was further supported by the HMBC spectrum of I . The aldehyde proton at δ 9.49 (-CHO) correlated with the carbons at δ 73. 1 (C-16), 51. 5 (C-17), and 40.8 (C-18). In the 13 CNMR spectrum of I , the signals of C-3 and C-23 were shifted downfield to δ 85. 6 and 78. 3 respectively as compared to signals of its aglycone I a. According to the rule of glycosylation shift, C-3 and C-23 of I must link with a monosaccharide, respectively. The HNM R spectrum of I had two anomeric proton signals at δ 5. 27 (d, \(\) 6. 8 Hz) and 5. 17 (d, \(\) 7. 8 Hz) and its 13 CNMR spectral data indicated the presence of terminal β-D-glucopyranosyl unit and α-D-ribofuranosyl unit^[4]. Based on the HMBC spectrum of I , the proton at δ 4.01 (H-3) had correlation with C-1 (δ 105.4) of glucose and the hydroxymethyl proton at δ 3.87 (H-23) had a cross-peak with C-1 (δ 103.7) of ribose. It was confirmed that glucose was attached to the C-3 position and ribose to C-23. Thus, the structure of I was elucidated to be 3, 16 dihydroxy-olean-12-en-28-al-3-O + D-glucopyranosyl-23-O + D-ribofuranoside. The key HMBC correlations for I (see Fig. 1), chemical shifts of I and I a (see Table 1).

2 Experimental

- 2.1 General experimental procedures IR spectrum was measured on a Nicolet MX-1 spectrometer as a pressed KBr disk. NMR spectra were recorded on Bruker AP-300 and DRX-500 MHz spectrum was TMS as the internal standard. MS spectra were measured on a VG AutoSpec-3000 mass spectrometer. Silica gel with 200-300 mesh was used for column chromatography.
- 2.2 Plant material The whole plant of L. candi

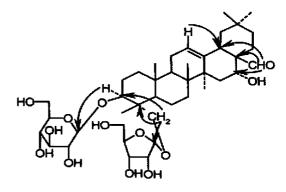


Fig. 1 The key HMBC correlations for ITable 1 Chemical shifts of compounds I (in $C_5 D_5 N$,

500 MHz for 1 H and 125 MHz for 13 C) and I a (in $C_5 D_5 N$, 75 MHz) (J Hz)

	1 a (11	1 C ₅ D ₅ N,	/5 MHZ) (J	111Z)	
С, Н	$^{13}{\rm CNM}~{\rm R}~{\rm of}$	13 CNM R of	¹ HNM R o	of	HM BC
atom	I *	I a	I		(H to C)
1	38. 9 (t)	38. 2			
2	27. 3 (t)	26. 2			
3	85. 6 (d)	78. 1	4. 01, dd	G	1, 5, Glc-1
4	37. 2 (s)	41.8			
5	51. 6 (d)	49.8			
6	17. 7 (t)	18.4			
7	32. 7 (t)	32. 7			
8	40. 0 (s)	39.7			
9	47. 0 (d)	46. 6			
10	37. 1 (s)	36. 8			
11	23. 6 (t)	23. 1			
12	123. 6 (d)	123.7	5. 45, brs	G	18
13	143. 6 (s)	142. 1			
14	40. 9 (s)	41.4			
15	35. 5 (t)	34. 6			
16	73. 1 (d)	73.0			
17	51. 5 (s)	50.7			
18	40. 8 (d)	40. 3			
19	46. 9 (t)	46. 2			
20	30. 8 (s)	30. 4			
21	34. 9 (t)	35.4			
22	23. 6 (t)	23. 3			
23	78. 3 (t)	73.5	3. 88, d, J= 1	10. 3 G	3, 4, Rib-1
			3. 86, d, J= 1	10. 3	
24	13. 8 (q)	11.4	1. 06, s	G	3, 4, 5, 23
25	16. 6 (q)	17. 1	0. 797, _s	\mathbf{G}	1, 5, 9, 10
26	17. 4 (q)	17.6	0. 739, s	G	7, 8, 9, 14
27	27. 1 (q)	26. 9	1. 65, s	G	8, 13, 14, 15
28	205. 7 (d)	204. 7	9. 49, _s	G	16, 17, 18
29	33. 3 (q)	32. 9	0. 977, _s	\mathbf{G}	19, 20, 21
30	24. 2 (q)	23. 9	1. 00, s	G	19, 20, 21

 $^{^*}$ Sugar moieties of I : Glc 105. 4 (d), 75. 2 (d), 78. 5 (d), 70. 5 (d), 78. 4 (d), 62. 7 (t); Rib 103. 7 (d), 71. 2 (d), 71. 7 (d), 83. 8 (d), 64. 4 (t)

da was collected in Mianyang, Sichuan Province, China and identified by Professor Xiao Shunchang, Chengdu Institute of Biology, Chinese Academy of Sciences. 2.3 Extraction and isolation The dried powder (7.25 kg) was extracted three times with MeOH for ten days each time at room temperature. After removal of the solvent, the residue was suspended in HeO and successively extracted with petroleum ether (bp 60 °C ~ 90 °C), EtOAc and n-BuOH. The n-BuOH extracts (108 g) were subjected to silica gel (CHClb-MeOH= 35: 1) column chromatography to obtain six fractions. Fraction sixth was chromatographed over silica gel eluted with CHClb-MeOH (20: 1) to afford compound I (30 mg).

2.4 Identification Compound I white powder, IRV $_{\rm max}^{\rm KBr}$ (cm $^{-1}$): 3.432, 2.946, 2.936, 1.710, 1.112, 1.075, 1.040. HR-FAB-MS [M + H † m/ 2 767, 455 8 (G41 H67 O15, calcd. 767, 458 2). FAB-MS (-) m/z 765 [M - H † , 603 [M - glc † . FABMS (+) m/z 767 [M+ H † , 605 [M - glc+ H † , 455 [M - glc- rib- H2 O+ H † .

Hydrolysis of I: Compound I (8 mg) hydrolyzed with 0.5 mol/L H_2SO_4 (5 mL, EtO H- H_2O = 1: 1) in boiling water bath for 4 h. After removal of EtOH under reduced pressure, the mixture was extracted with chloroform three times. The organic layer was evaporated to dryness to give I a (5 mg).

Acetylation of I: Compound I (2 mg) was acetylated with A&O-pyridine (1:2,0.75 mL) at room temperature for 48 hours to yield acetate of I. EI-MS (m/z): 331,259.

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山楂化学成分研究

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关键词: 山里红;牡荆素鼠李糖苷;金丝桃苷

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Studies on chemical constituents from fruit of Crataegus pinnatifida

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Abstract Object To look for the proprietary constituent and the constituents with blood lipid regulating effect from the dried fruit of *Crataegus pinnatifida* Bge. var. *major* N. E. Br. **Methods** Various column chromatographic techniques were employed for isolation and purification of the constituents. UV,

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