

· Reviews ·

Recent Advance in Chemical and Biological Studies on Cimicifugeae Pharmaceutical Resources

HAO Da-cheng^{1*}, GU Xiao-jie¹, XIAO Pei-gen^{2*}, LIANG Zhan-guo¹, XU Li-jia², PENG Yong²

1. Biotechnology Institute, School of Environmental and Chemical Engineering, Dalian Jiaotong University, Dalian 116028, China

2. Key Laboratory of Bioactive Substances and Resources Utilization of Chinese Herbal Medicine, Ministry of Education, Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences, Beijing 100193, China

Abstract: Cimicifugeae is one of the rich sources for various active components and the health promoting and therapeutic values of the components have been corroborated by long-term use in folk medicine and traditional Chinese medicine. Increasing interest in Cimicifugeae pharmaceutical resources has led to the further discoveries of triterpenoid saponins, phenolic compounds, chromones, and many other compounds in various species of Cimicifugeae, and to the investigations on their chemotaxonomy, molecular phylogeny, and bioactivities. Based on our pharmacophylogenetic studies, the progress in phytochemistry, chemotaxonomy, molecular biology, and phylogeny of Cimicifugeae had been summarized since 2007, especially *Cimicifuga* L. ex Wernisch. and *Actaea* L., and their relevance to therapeutic efficacy. An exhaustive literature survey is used to characterize the global scientific effort in the phytochemical and biological studies of Cimicifugeae. More triterpenoid saponins have been found in various species, among which the cimigenol type (type A) is predominant. The versatile bioactivities of saponins and extracts, as well as those of phenolics and other ingredients, were summarized and discussed. The morphology-based five-genus classification of Cimicifugeae is not supported by molecular phylogeny. Molecular phylogeny based on nuclear and chloroplast DNA sequences tends to merge *Cimicifuga* Wernisch., *Souliea* Franch., and *Actaea* L. into a single genus. It is indispensable to integrate the emerging technologies into Cimicifugeae research for both the sustainable utilization of Cimicifugeae pharmaceutical resources and finding novel compounds with potential clinical utility and less adverse effects. Systems biology and omics technologies would play an increasingly important role in booming pharmaceutical research involving bioactive compounds of Cimicifugeae.

Key words: biological activity; chemotaxonomy; Cimicifugeae; pharmaceutical resource; phylogeny; phytochemistry

DOI: 10.3969/j.issn.1674-6348.2013.02.001

Introduction

Cimicifugeae plants, belonging to subfam. Helleboroideae and Ranunculaceae, are traditionally composed of five genera *Cimicifuga* L. ex Wernisch., *Actaea* L., *Anemonopsis* Siebold et Zucc., *Souliea* Franch., and *Beesia* Balf. f. et W. W. Sm. There are more than 40 species in the tribe Cimicifugeae, which are native to temperate regions of the Northern Hemisphere. The genus *Actaea* L. is closely related to

Cimicifuga L. ex Wernisch. and *Souliea* Franch., and based on combined DNA sequence data and similarity in biochemical constituents and morphology, many botanists included those two genera within *Actaea* L. (Compton and Culham, 2002). If not merged, *Cimicifuga* L. ex Wernisch. has 28 species and eight are distributed in China. Among them, *Cimicifuga foetida* L., *C. dahurica* Maxim., and *C. heracleifolia* Komar. are officially listed in *China Pharmacopoeia 2010* as

* Correspond authors: Hao DC E-mail: hao@djtu.edu.cn

Xiao PG E-mail: xiaopg@public.bta.net.cn

Receive: October 10, 2012; Revised: January 15, 2013; Accepted: March 1, 2013

Fund: Dalian Jiaotong University and Key Laboratory of Bioactive Substances and Resources Utilization of Chinese Herbal Medicine, Ministry of Education, Ministry of Science and Technology national support program (2012BAI29B01)

Online time: April 28, 2013 Online website: <http://www.cnki.net/kcms/detail/12.11410.R.20130428.0911.003.html>

“*Shengma*”, which have been used since ancient times to treat wind-heat headache, sore throat, toothache, and uterine prolapse, etc. Other species of *Cimicifuga* L. ex Wernisch. are also used in folk medicine to treat physical wounds, promote eruption, and detoxify (Gao *et al.*, 2008). There are eight species in genus *Actaea* L., and six of them are distributed in North America. Among them, *Cimicifuga racemosa* (Nutt.) L. (CR), also called black cohosh, has long been used by native American to tackle rheumatism, menopausal, and nervous problems. Presently, black cohosh products are among the most popular health-promoting medicines in the US and Europe (Jiang *et al.*, 2006). Two species of *Actaea* L. are used in traditional Chinese medicine and their therapeutic use is distinct from *Cimicifuga* Wernisch. (Gao *et al.*, 2008). *Souliea* medicine is antipyretic and antitoxic, which is also used to eliminate dampness, clear heart fire, and relieve anxiety. *Beesia* Balf. f. et W. W. Sm. is endemic in China and is used to treat cold caused by exterior heat, joint pain of rheumatism, diarrhea, sore throat, headache, and snake bite. On the phylogenetic tree based on chloroplast and

nuclear DNA sequences, *Beesia* Balf. f. et W. W. Smith. and the monotypic genus *Anemonopsis* Siebold et Zucc. together, and are basal to the cluster formed by *Cimicifuga* L. ex Wernisch., *Actaea* L., and *Souliea* Franch. (Wang *et al.*, 2009). However, little is known about the chemical constituents and bioactivities of these two genera. *C. foetida* and *A. racemosa* are the most frequently studied. In this brief review, we summarized the recent progress in the phytochemical and biological studies of Cimicifugeae since 2007.

Triterpenoid saponins

The structures of rings A—D are not quite different in diverse cycloartane type saponins, whereas the side chains tied with ring D vary greatly, due to the difference in oxygenation and the approach of ring formation (Table 1 and Fig. 1).

Four new cycloartane compounds are structurally peculiar, namely, there is C-C bond cleavage between C-15 and C-16, as well as the six-membered lactone ring between C-15 and C-23 (Nian *et al.*, 2012; Yoshimitsu, Nishida, and Nohara, 2007). Ring D of all

Table 1 Triterpenoid saponins found in species of Cimicifugeae since 2007

No.	Compounds	Aglycone	Species	Tissue	References
1	isocimipodocarpaside	L	<i>Cimicifuga racemosa</i>		Jamróz <i>et al.</i> , 2012
2	3β,16α-dihydroxy-12-acetoxy-16,22-cyclo-23-ketone-24R,25-epoxy-cycloartane-7-ene 3-O-β-D-galactopyranoside	R	<i>C. simplex</i>		Kuang <i>et al.</i> , 2012
3	24-O-hydroxy-7,8-didehydrohydroshengmanol 3-O-β-D-galactopyranoside	B			
4	24- <i>epi</i> -24-O-hydroxy-7,8-didehydrohydroshengmanol 3-O-β-D-galactopyranoside	B			
5	methyl 3,4-seco-4-hydroxy-3-cimigenolate	A	<i>C. foetida</i>	aerial parts	Nian <i>et al.</i> , 2012
6	cimigenol-3-O-[2',4'-O-diacetyl]-α-L-arabinopyranoside	A			
7	cimigenol-3-O-[3',4'-O-diacetyl]-α-L-arabinopyranoside	A			
8	cimigenol-3-O-[4'-O-acetyl]-α-L-arabinopyranoside	A			
9	25-anhydrocimigenol-3-O-[3'-O-acetyl]-α-L-arabinopyranoside	A			
10	24- <i>epi</i> -cimigenol-3-one	A			
11	15,16- <i>seco</i> -7,8-didehydro-15-formyl-16-oxohydroshengmanol	P			
12	7,8-dihydro-11-dehydroxycimicidanol	D			
13	shengmanol-3-O-[2'-O-acetyl]-α-L-arabinopyranoside	B			
14	24- <i>epi</i> -cimigenol-3-one	A	<i>C. foetida</i>		Lu <i>et al.</i> , 2012
15	foetinoside	M			
16	cimipodocarpaside	L	<i>C. racemosa</i>		Jamróz <i>et al.</i> , 2011
17	shengmaxinsides A	A	<i>C. simplex</i>	root	Kuang <i>et al.</i> , 2011
18	shengmaxinsides B	A			
19	shengmaxinsides C	B			
20	23-O-methyl-24-deoxy-2'-O-(3''-methylmalonyl)-cimiaceroside B	E	<i>C. foetida</i>	rhizomes	Sun <i>et al.</i> , 2011

(To be continued)

(Continued Table 1)

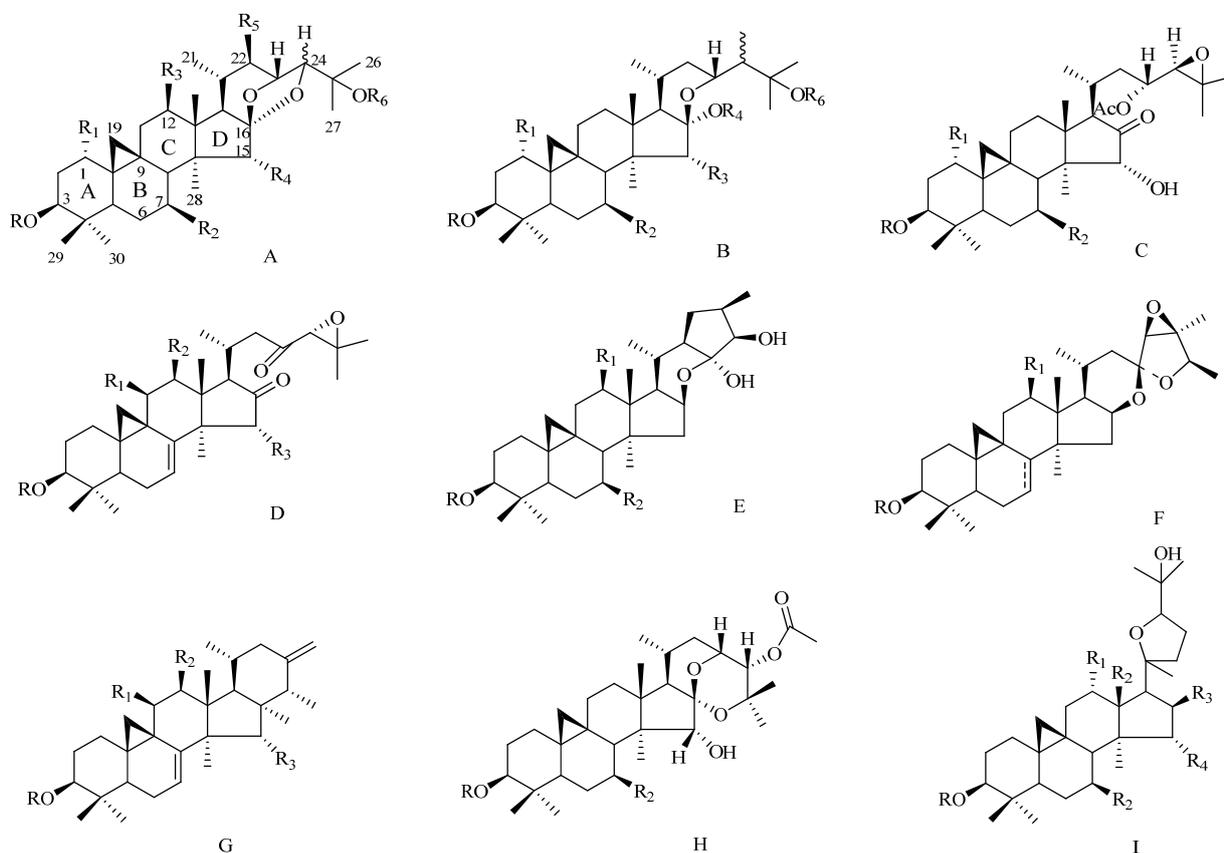
No.	Compounds	Aglycone	Species	Tissue	References
21	2'- <i>O</i> -(3''-methylmalonyl)actein	F			
22	2',24-di- <i>O</i> -acetylisodahurinol-3- <i>O</i> - α - <i>L</i> -arabinopyranoside	B	<i>C. foetida</i>	aerial parts	Nian <i>et al</i> , 2011
23	24- <i>O</i> -acetylisodahurinol-3- <i>O</i> - α - <i>L</i> -arabinopyranoside	B			
24	12 β -hydroxy-25-anhydrocimigenol	A			
25	cimigenol-12-one	A			
26	12 β -hydroxy-15-deoxycimigenol	A			
27	2'- <i>O</i> -acetyl-24-epi-cimigenol-3- <i>O</i> - α - <i>L</i> -arabinopyranoside	A			
28	2'- <i>O</i> -acetylcimigenol-3- <i>O</i> - β - <i>D</i> -xylopyranoside	A			
29	25-anhydrocimigenol-3- <i>O</i> - α - <i>L</i> -arabinopyranoside	A			
30	2',23-di- <i>O</i> -acetylshengmanol-3- <i>O</i> - α - <i>L</i> -arabinopyranoside	C			
31	2',24-di- <i>O</i> -acetyl-25-anhydroshengmanol-3- <i>O</i> - α - <i>L</i> -arabinopyranoside	B			
32	3 β ,15 α ,16 α ,24 α -tetrahydroxy-25,26,27-trinor-16,24-cyclo-cycloartane-23-one 3- <i>O</i> - β - <i>D</i> -xylopyranoside	G	<i>C. heracleifolia</i>	rhizomes	Nishida and Yoshimitsu, 2011
33	3 β ,15 α ,16 α ,24 α -tetrahydroxy-25,26,27-trinor-16,24-cyclo-cycloart-7-en-23-one 3- <i>O</i> - β - <i>D</i> -xylopyranoside	G			
34	12 β -acetoxy-3 β ,15 α ,16 α ,24 α -tetrahydroxy-25,26,27-trinor-16,24-cyclo-cycloart-7-en-23-one 3- <i>O</i> - β - <i>D</i> -xylopyranoside	G			
35	3 β ,11 β -dihydroxy-24,25,26,27-tetranor-cycloart-7-en-23,16 β -olide 3- <i>O</i> - β - <i>D</i> -xylopyranoside	O			
36	23 <i>R</i> ,24 <i>S</i> -diacetoxy-3 β ,15 α ,25-trihydroxy-cycloart-7-en-16-one 3- <i>O</i> - β - <i>D</i> -xylopyranoside	M			
37	23 <i>R</i> -acetoxy-3 β ,15 α ,24 <i>R</i> ,25-tetrahydroxy-cycloart-7-en-16-one 3- <i>O</i> - β - <i>D</i> -xylopyranoside	M			
38	foetidinosides A	S	<i>C. foetida</i>		Lu <i>et al</i> , 2010
39	foetidinosides B	L			
40	foetidinosides C	M			
41	foetidinosides D	M			
42	foetidinosides E	M			
43	3- <i>O</i> - β - <i>D</i> -xylopyranosyl cimigenol 15- <i>O</i> - β - <i>D</i> -glucopyranoside	A	<i>C. foetida</i>	rhizomes	Shen, 2010
44	25- <i>O</i> -acetyl cimigenol 3- <i>O</i> - β - <i>D</i> -xylopyranosyl 15- <i>O</i> - β - <i>D</i> -galactopyranoside	A			
45	cimifoetiside A		<i>C. foetida</i>	aerial parts	Pan <i>et al</i> , 2009; 2009b
46	cimifoetiside B				
47	15 α -hydroxy-16-dehydroxy-16(24)-en-foetidinol-3- <i>O</i> - β - <i>D</i> -xylopyranoside	G	<i>C. foetida</i>	rhizomes	Lu <i>et al</i> , 2009
48	28-hydroxy-foetidinol-3- <i>O</i> - β - <i>D</i> -xylopyranoside	G			
49	foetidinol-3- <i>O</i> - β - <i>D</i> -xylopyranosyl-(1'' \rightarrow 3')- β - <i>D</i> -xylopyranoside	G			
50	(3',12 β)- <i>O</i> -diacetyl-cimigenol-3- <i>O</i> - β - <i>D</i> -xylopyranoside	A	<i>Actaea asiatica</i>	rhizomes	Fan <i>et al</i> , 2009
51	(4',25)- <i>O</i> -diacetyl-cimigenol-3- <i>O</i> - β - <i>D</i> -xylopyranoside	A			
52	2'- <i>O</i> -acetyl-25- <i>O</i> -methyl-cimigenol-3- <i>O</i> - β - <i>D</i> -xylopyranoside	A			
53	2'- <i>O</i> -acetyl-25- <i>O</i> -ethyl-cimigenol-3- <i>O</i> - β - <i>D</i> -xylopyranoside	A			
54	3'- <i>O</i> -acetyl-cimicifugoside	F			
55	4'- <i>O</i> -acetyl-23-epi-26-deoxycimicifugoside	F			
56	(23 <i>R</i>)-26-deoxycimicifugoside	F	<i>A. asiatica</i>	rhizomes	Lu, Fan, and Duan, 2008
57	cimiaceroside C	E	<i>C. foetida</i>	rhizomes	Sun <i>et al</i> , 2008
58	cimifosides A	A			
59	cimifosides B	A			
60	cimifosides C	B			
61	cimifosides D	F			
62	7,8-dihydroactaeapoxide 3- <i>O</i> - β - <i>D</i> -xylopyranoside	N	<i>A. pachypoda</i>	roots	Ali <i>et al</i> , 2007b

(To be continued)

(Continued Table 1)

No.	Compounds	Aglycone	Species	Tissue	References
63	12-deacetoxyactaeaepoxide 3- <i>O</i> - β - <i>D</i> -xylopyranoside	N			
64	12 β -acetoxycimigenol	A			
65	podocarpasides A	L	<i>A. podocarpa</i>	roots	Ali <i>et al</i> , 2007a
66	podocarpasides B	L			
67	podocarpasides C	L			
68	podocarpasides D	L			
69	podocarpasides E	L			
70	podocarpasides F	L			
71	podocarpasides G	L			
72	podocarpasides H	D	<i>A. podocarpa</i>	roots	Ali, Khan, and Khan, 2007
73	podocarpasides I	D			
74	podocarpasides J	D			
75	24-acetoxy-15,16- <i>seco</i> -cycloartane 3- <i>O</i> - β - <i>D</i> -xylopyranoside	P	<i>Cimicifuga</i>	rhizomes	Yoshimitsu, Nishida, and Nohara, 2007
76	24- <i>epi</i> -hydro-15,16- <i>seco</i> -cycloartane 3- <i>O</i> - β - <i>D</i> -xylopyranoside	P			
77	24- <i>epimer</i> -hydro-15,16- <i>seco</i> -cycloartane 3- <i>O</i> - β - <i>D</i> -xylopyranoside	P			
78	cimifoetiside VI	B	<i>C. foetida</i>	aerial parts	Pan <i>et al</i> , 2007
79	cimifoetiside VII	B			
80	cimicifugadine	Q	<i>C. foetida</i>	roots	Dan <i>et al</i> , 2007
81	cimicifoetisides A	A	<i>C. foetida</i>	rhizomes	Sun <i>et al</i> , 2007
82	cimicifoetisides B	A			

A: cimigenol type B: hydroshengmanol type C: shengmanol type D: 16,23-dione type E: cimiacerogenin type F: cimifugenin type G: side chain-*seco* type H: neocimicigenoside type I: beesioside type J: asiaticoside type K: ranunculane type L: 9,10-*seco*-9,19-cycloartane type M: 9,19-cycloartane type N: actaeaepoxide type O: tetranor type P: 15,16-*seco*-cycloartane type Q: cycloartane triterpene alkaloid type R: 16,22-*seco*-cycloartane type S: lanostane type; same as below



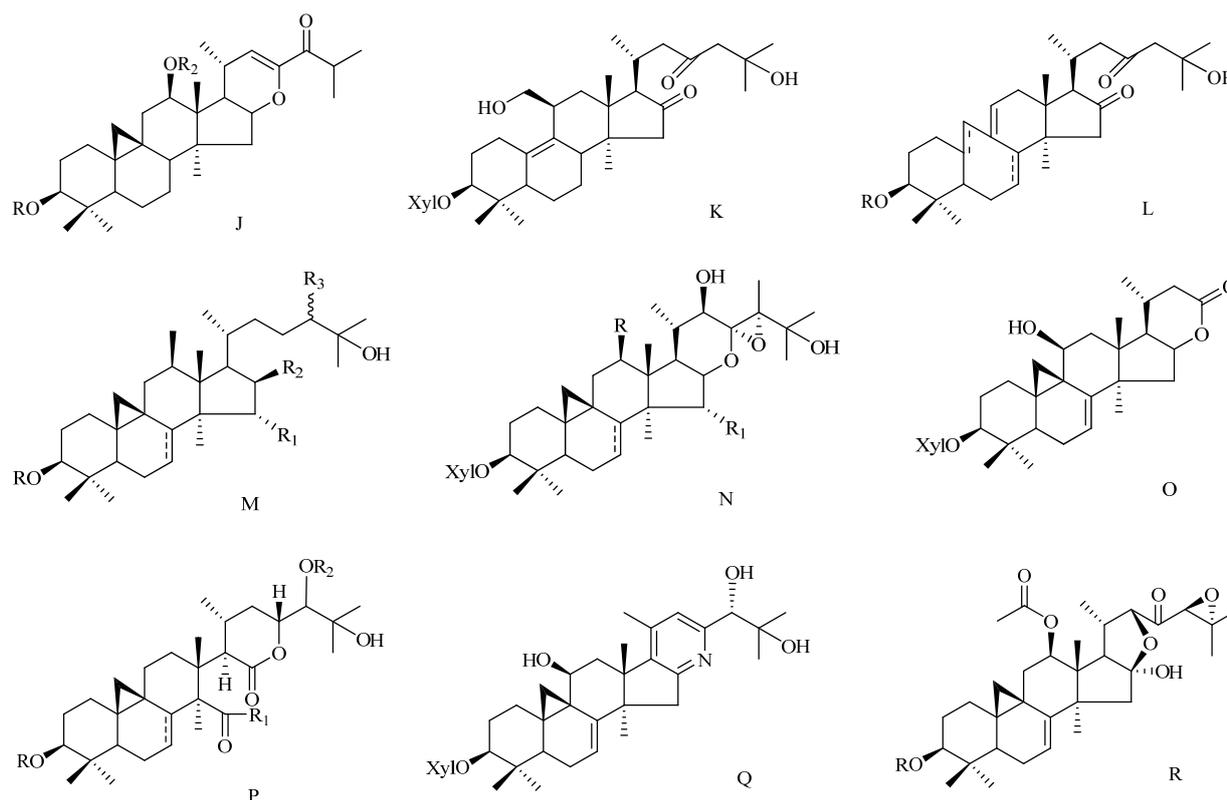


Fig. 1 Chemical structure types of 9,19-cycloartane triterpenoids in tribe Cimicifugeae

other cycloartane compounds is closed. An unprecedented triterpene alkaloid glycoside, designated cimicifugadine, with a pyridine ring incorporated into a cycloartane triterpenoid nucleus, has been isolated from *C. foetida* (Dan *et al.*, 2007). Several glycosides are characterized as 9,10-*seco*-9,19-cycloartane type with change in the cleavage between C-9 and C-10 (Jamróz *et al.*, 2012; 2011; Lu *et al.*, 2010; Ali *et al.*, 2007a). C-3 β is frequently glycosylated and usually linked with one or more sugars. The most common sugar is pentose (usually xylose and arabinose), while hexose (e.g., glucose and galactose) is occasional. Sometimes C-15, 16, 22, 24, and 25 are also glycosylated.

Bioactivities and adverse effects of saponins and extracts from Cimicifugeae

Anticancer activity

Novel anticancer compounds from natural product, due to their safety and efficiency, provide a gorgeous substitute to synthetic chemicals. Cimigenol from *C. foetida* exerted potent cytotoxic activity against SMMC-7721 (7.87 $\mu\text{mol/L}$) and A-549 (12.16 $\mu\text{mol/L}$), while cimiacerin B also showed the obvious

cytotoxicity against A-549 cell line (Lu *et al.*, 2012). The research group led by Prof. XIAO Pei-gen found that *C. foetida* extract inhibited the proliferation of hepatocellular cells via the induction of cell cycle arrest and apoptosis (Tian *et al.*, 2007a). CR (black cohosh) reduced Ki-67 and cyclin D1 protein expression in fibroadenoma and may have the chemopreventive potential for breast cancer (Einbond *et al.*, 2012). Cimicide E from *C. heracleifolia*, one of source plants of *C. foetida*, arrests cell cycle and induces the cell apoptosis in gastric cancer cells (Guo *et al.*, 2009). Total glycosides from the aerial parts of *C. dahurica* Thurez Maxim. (TGC), another source plant of *C. foetida*, induced G₀/G₁ HepG2 cell cycle arrest at lower concentration and triggered G₂/M arrest and apoptosis at higher concentration (Tian *et al.*, 2007b). An increase in the ratio of Bax/Bcl-2 was implicated in TGC-induced apoptosis. Besides, TGC dose-dependently inhibited the growth of the implanted H₂₂ tumor in mouse, treatment with the triterpene glycoside actein induced a stress response and apoptosis in human breast cancer cells, suggesting that compounds from the species of *Cimicifuga* L. ex Wernisch. may be useful in

the prevention and treatment of breast cancer (Einbond *et al.*, 2008). Gene expression profiling reveals the effects of CR on the estrogen receptor positive breast cancer cell line MCF-7 (Gaubé *et al.*, 2007). No estrogenic but antiproliferative and proapoptotic gene expression was shown for CR in MCF-7 cells at the transcriptional level. The effects may be caused by the activation of different pathways. The cycloartane glycosides and their aglycones could be identified as the active components in CR. The methanolic extract of CR also activated genes that enhanced apoptosis and repressed cell cycle genes in MDA-MB-453 human breast cancer cells (Einbond *et al.*, 2007). The CR extract BNO 1055 inhibited the proliferation of human prostate cancer-derived LNCaP cells (Seidlová-Wuttke, Thelen, and Wuttke, 2006).

Three cycloartane triterpenoids from the aerial parts of *C. foetida* exhibited broad-spectrum and moderated cytotoxic activities (Nian *et al.*, 2011). Human heat shock protein (Hsp) 27 was increased in various human cancer cells and exhibited cytoprotective activity that could affect tumorigenesis and the susceptibility of tumors to cancer therapy, but its expression at 2000 mg/L *C. foetida* extract was diminished (Soler *et al.*, 2011). Three 9,19-cycloartane triterpene glycosides exhibited the significant cytotoxicity against human HepG2 cells (Nian *et al.*, 2010). Cimicifugoside, a triterpenoid from *C. simplex*, is a specific nucleoside transport inhibitor that could display the potentiation of methotrexate cytotoxicity (Yawata *et al.*, 2009). The triterpene glycoside actein activated stress- and statin-associated responses and was bioavailable in Sprague-Dawley rats (Einbond *et al.*, 2009). Actein reduced free fatty acid and cholesterol content in the liver and inhibited the growth of HepG2 liver cancer cells.

Anemopsis californica (Nutt.) Hook. & Arn. showed *in vitro* anticancer activity against human colon cancer cells HCT-8 and the breast cancer cells (Kaminski *et al.*, 2010). The growth inhibitory effect of *A. californica* in breast cancer cells was extracellular signal-regulated kinase (ERK)-mediated (Daniels *et al.*, 2006). 25-Anhydrocimigenol-3-*O*- β -*D*-xylopyranoside isolated from *Souliea vaginata* (Maxim.) Franch. showed the anticancer activity against hepatoma, as it might induce apoptosis and G₀/G₁ cell cycle arrest

(Tian *et al.*, 2006). The anticancer activity of *Beesia Balf. f. et W. W. Sm.* awaits further studies. In the future, the structure-activity relationship of various saponins and their potentials in the treatment of various human cancers should be studied for novel drug development.

Effects on menopausal symptoms

CR has been used in Europe as a medicinal plant for more than a century and its roots have been widely used for the treatment of menopausal symptoms. Recently randomized studies have shown that CR consumption alleviated “hot flush” and due to the lack of uterotrophic effects it could be a safe option for estrogen replacement therapy (Rachoń *et al.*, 2008). The isopropanol extract of CR may act on the hypothalamic nuclei and have the therapeutic effects on menopausal symptoms (Zhang *et al.*, 2012). Hot flush is a disorder of thermoregulation due to the lack of estrogens and is the most common and characteristic climacteric complaint. The CR special extract BNO 1055 could prevent hot flashes in ovariectomized rats (Kapur, Wuttke, and Seidlova-Wuttke, 2010). The combination of CR with *Hypericum perforatum* L. (HP) had a positive effect on climacteric complaints, while CR monotherapy as well as HP and *Vitex agnus-castus* L. was not better than placebo (Laakmann *et al.*, 2012). A standardized isopropanolic CR extract (remifemin) was safe and effective for menopausal symptoms (Ross, 2012). It was an effective agent to manage Chinese women with climacteric symptom (Bai *et al.*, 2009). It had the similar therapeutic effect and lower incidence of adverse effect when compared with Tibolone. For the CR extract, the onset of affecting abnormal thermoregulation took longer than that of estradiol valerate in ovariectomized rats (Ma *et al.*, 2011b). CR extract had a significant effect on day central body temperature (CBT) but did not affect night CBT of ovariectomized rats. CR extract might be a reasonable treatment in tamoxifen-treated breast cancer patients with predominantly psychovegetative symptoms (Rostock *et al.*, 2011). It was hypothesized that the established positive allosteric modulation of γ -aminobutyric acid (GABA) type A receptors might contribute to the beneficial effects of CR extracts in the treatment of climacteric symptoms (Cicek *et al.*, 2010). Four cycloartane glycosides of CR significantly enhanced

GABA-induced chloride currents.

Osteoprotective effects

Triterpenoids from *Cimicifugae Rhizoma* are a novel class of inhibitors on bone resorption and ovariectomy-induced bone loss (Li *et al.*, 2007). CR extract had osteoprotective effects and its triterpene saponins were responsible for the reduction of bone marrow fat and the secretion of pro-inflammatory cytokines (Seidlova-Wuttke *et al.*, 2012a; 2012b). CR facilitated metaphyseal fracture healing in the early stage of osteoporosis in ovariectomized rats (Kolios *et al.*, 2010). Deoxyactein isolated from CR protected osteoblastic MC3T3-E1 cells against antimycin A-induced cytotoxicity (Choi, 2011). Deoxyactein stimulated osteoblast function and inhibited bone-resorbing mediators in MC3T3-E1 cells (Choi, 2013). *C. heracleifolia* significantly preserved trabecular bone mass, bone volume, trabecular number, trabecular thickness, structure model index, and bone mineral density of proximal tibia metaphysis or distal femur metaphysis, thus preventing ovariectomy-induced bone loss in mice (Ahn *et al.*, 2012). Cycloartane triterpenoids of *C. foetida*, such as cimigenol, actein, (23*R*,24*S*)25-*O*-acetyl-cimigenol-3-*O*- β -*D*-xylopyranoside, and (23*R*,24*S*)cimigenol-3-*O*- β -*D*-xylopyranoside, exhibited osteoclast inhibition activity (Dan *et al.*, 2009). A triterpene glycoside from CR inhibited osteoclastogenesis by modulating tumor necrosis factor (TNF)- α and RANKL (a member of the TNF superfamily) signaling pathways (Qiu *et al.*, 2007). The special extract of CR BNO 1055 was shown to have bone protective effects without exerting estrogenic effects in the uterus or mammary gland, as well as other organs that also express estrogen receptors (Seidlova-Wuttke, Jarry, and Wuttke, 2009).

Anti-inflammatory and immunosuppressive activities

Cycloartane-type triterpene glycosides from the rhizomes of *C. heracleifolia* and *C. foetida* showed anticomplement activity (Lee *et al.*, 2012; Qiu *et al.*, 2006). Cyclolanostane triterpene diglycosides from the aerial part of *C. foetida* effectively inhibited the proliferation of murine splenocytes induced by concanavalin A (Pan *et al.*, 2009). A non-physiological accumulation of fat cells in abdomen and joints could result in the increased production of proinflammatory

cytokines that have the adverse effects on serum lipids and glucose and on joint cartilage. The special extract of CR BNO 1055 was shown to reduce the size of the fat depot. This extract and its saponin prevented the metabolic syndrome and deterioration of cartilage in the knee joint of ovariectomized rats (Seidlova-Wuttke *et al.*, 2012a; 2012b). Combined prescription of *Aralia cordata* Thunb. and *C. heracleifolia* and its major compounds inhibited matrix proteinases and vascular endothelial growth factor through the regulation of mitogen-activated protein kinase (MAPK) pathway, thus providing a therapy for osteoarthritis (Huh *et al.*, 2011). Isoimperatorin, cimicide E, and 23-*O*-acetylshengmanol-3-xyloside from *Cimicifugae Rhizoma* inhibited TNF- α -induced vascular cell adhesion molecule (VCAM)-1 expression in human endothelial cells (Moon *et al.*, 2011). Peroxisome proliferator-activated receptor- γ (PPAR- γ) upregulation and PI3K, ERK 1/2, and protein kinase C (PKC) signal pathways are involved in their anti-inflammatory activities. Cimicemate A from multiple species of *Cimicifuga* L. ex Wernisch. suppressed the lipopolysaccharide-induced TNF α production in the blood macrophages (Yang *et al.*, 2009). It might modulate the activities of signaling MAPK and transcription factor such as nuclear factor- κ B (NF- κ B).

Effects on cardiovascular system

CR relaxed the isolated rat thoracic aorta through endothelium-dependent and -independent mechanisms (Kim, Lee, and Rhyu, 2011). CR extract elicited the vasorelaxant effect via the NO/cGMP pathway. CR-induced endothelium-independent vasorelaxation appeared to involve the inhibition of calcium influx mediated by the opening of inward rectifier potassium channels. 7,8-Didehydrocimigenol (DHC), a triterpenoid of *Cimicifugae Rhizoma*, could increase the expression of PPAR- γ in endothelial cells (ECs) in a time- and dose-dependent manner (Mun *et al.*, 2011). 7,8-DHC could inhibit TNF- α -induced expression of VCAM-1 but not ICAM-1 through upregulation of PPAR- γ in human ECs and could be used for the treatment of cardiovascular disorders such as atherosclerosis.

Other positive effects

Ig administration of CR extract attenuated psychological and physiological stress responses (Nadaoka *et al.*, 2012a). The CR extract could alleviate

the acute stress responses by suppressing the changes of amine-to-metabolite ratio in brain (Nadaoka *et al.*, 2012b). The CR extract significantly prevented the development of water immersion stress-induced gastric mucosal ulcers in rats (Nadaoka *et al.*, 2012c).

CR extracts reduced endometrial proliferation in comparison to the placebo (Alves *et al.*, 2008). CR decreased local estrogen formation in normal human breast tissue *in vitro*, which might contribute to the lack of hormonal effects of black cohosh in breast tissue observed in previous studies (Stute *et al.*, 2007). Adding dry extract from CR rhizome to clomiphene citrate induction could improve the pregnancy rate and cycle outcomes in couples with unexplained infertility and recurrent clomiphene citrate induction failure (Shahin *et al.*, 2008).

Dichloromethane fraction of *C. heracleifolia* decreased the level of melanin synthesis by activating the ERK or AKT signaling pathway in B16F10 cells (Jang *et al.*, 2009). *C. heracleifolia* would be a useful therapeutic herb for treating hyperpigmentation and an effective component in whitening and/or lightening cosmetics.

Adverse effects and safety

Eight triterpene glycosides of CR were identified as competitive CYP (cytochrome P450) 3A4 inhibitors with IC₅₀ values ranging from 2.3–5.1 μmol/L, while the alkaloids protopine and allocryptopine were identified as competitive CYP2D6 inhibitors (Li *et al.*, 2011). Co-administration of CR with tamoxifen might interfere with the clinical efficacy of this drug. CYPs 1A2, 2D6, 2C9, 2C19, and 3A4 were inhibited *in vitro* by CR extracts (Huang *et al.*, 2010; Ho *et al.*, 2011). Fukinolic acid derivatives and triterpene glycosides from CR inhibited CYP isozymes, but were not cytotoxic to HepG2 cells *in vitro*. However, Pang *et al.* (2011) suggested that the incidence of herb-drug interaction in patients administered with CR might not be mediated by human pregnane X receptor and CYP3A4. With the exceptions of St. John's wort and goldenseal, the currently available information suggested that concomitant intake of the herbal drugs, e.g., the well-known ginseng and CR, was not a major risk for drugs that were metabolized by CYPs (Zadayan and Fuhr, 2012).

CR extract induced dose-dependent hematological

changes, i.e., a non-regenerative macrocytic anemia, and increased the frequencies of peripheral micro-nucleated red blood cells in female B6C3F1/N mice and Wistar Han rats (Mercado-Feliciano *et al.*, 2012). CR would not influence breast cancer risk if given to women before tumor formation. However, it increased metastatic mammary cancer in transgenic mice expressing c-erbB2 (Davis *et al.*, 2008). CR impaired the fatty acid β-oxidation and induced oxidative stress in livers of ovariectomized rats with renovascular hypertension (Campos *et al.*, 2012). The use of CR might not exert an explicit hepatotoxicity risk, but quality problems in a few CR products were evident that required additional regulatory quality specifications (Teschke *et al.*, 2011).

Bioactivity of other compounds in tribe Cimicifugeae

Phenolic compounds

Petasiphenone, a phenol isolated from CR, inhibited the proliferation of the human prostate cancer cell line LNCaP *in vitro* (Jarry *et al.*, 2007). The radical scavenging activity of the extracts from *Actaea L.* correlated to their polyphenolic composition (Nuntanakorn *et al.*, 2007). Isoferulic acid (3-hydroxy-4-methoxycinnamic acid) from *Cimicifugae Rhizoma* is an effective natural anti-oxidant in both lipid and aqueous media (Wang, Li, and Chen, 2011). Actaealactone and a new phenylpropanoid ester derivative, cimicifugic acid G from CR, displayed the anti-oxidative activity in 1,1-diphenyl-2-picrylhydrazyl (DPPH) free-radical assay (Nuntanakorn *et al.*, 2006).

Fukinolic acid and cimicifugic acids A–J, from a mixture of *C. dahurica* and *C. heracleifolia*, showed stronger hyaluronidase inhibitory activities than the positive control, rosmarinic acid (Iwanaga *et al.*, 2010a). Cimicifugic acids K–N from *C. simplex* showed more potent hyaluronidase inhibitory activities than rosmarinic acid (Iwanaga *et al.*, 2010b). Isoferulic acid was the major active principle in CR root extract, responsible for the observed inhibition of interleukin (IL)-6, TNF-α, and interferon-γ, but not for IL-8 stimulation (Schmid *et al.*, 2009). The effect of this compound may explain the anti-inflammatory activities of CR and its beneficial actions in rheumatism and other inflammatory diseases.

Others

Cimicifugin from *C. foetida* inhibited human respiratory syncytial virus (HRSV; Wang *et al.*, 2012a). *C. foetida* dose-dependently inhibited viral attachment and could increase heparin effect on viral attachment (Wang *et al.*, 2012b). Additionally, *C. foetida* time- and dose-dependently inhibited HRSV internalization.

Serotonergic receptors and transporters are involved in thermoregulation. Various guanidine alkaloids and Pictet-Spengler adducts were detected in CR (Gödecke *et al.*, 2009). These strongly basic and frequently zwitterionic nitrogenous metabolites contribute considerable chemical diversity to the polar serotonergic fraction of CR, which is used to alleviate menopausal symptoms. New phytochemical methods and liquid chromatography-mass spectrometry (LC-MS) led to the identification of *N*(ω)-methylserotonin as serotonergic active principle of CR (Gödecke *et al.*, 2009). *N*(ω)-methylserotonin showed 5-hydroxytryptamine (serotonin) 7 receptor binding, induced cAMP and blocked serotonin re-uptake, suggesting that *N*(ω)-methylserotonin might be responsible for the serotonergic activity of CR (Powell *et al.*, 2008).

The oils extracted from the roots of *A. californica* demonstrated the antiproliferative activity against AN3CA and HeLa cells *in vitro* (Medina-Holguín *et al.*, 2008). Steam-distilled oil of *A. californica* had antimicrobial properties against three of 11 microbial species tested (Medina *et al.*, 2005). This bioactivity could be partially accounted by the α -pinene in oil. A new 4 α -methyl sterol from the aerial parts of *C. foetida*, cimisterol A, exhibited broad-spectrum and potent cytotoxic activities against human HL-60, Jurkat, K562, U937, HepG-2, and SGC-7091 cell lines (Nian *et al.*, 2012). The bioactivities of Cimicifugeae alkaloids await further studies.

Chemotaxonomy and authentication of Cimicifugeae

Black cohosh (*A. racemosa* and *C. racemosa*) is ranked among the top-selling herbs in the US. There is a risk for the adulteration with the similar-looking *C. americana*, which grows in the same habitats of the eastern US. Other adulterants found in the current global market are the Asian *Cimicifuga* L. ex Wernisch. species *C. foetida*, *C. heracleifolia*, and *C. dahurica*.

Three of the 11 black cohosh products were found to contain the marker compound—cimifugin but not cimracemoside C, indicating that these products contain Asian *Actaea* L. instead of black cohosh (Jiang *et al.*, 2006). One product contained both black cohosh and an Asian species of *Actaea* L. The chemotaxonomic distinctiveness of the HPLC fingerprints allowed the identification of ten species of *Cimicifuga* L. ex Wernisch., including three North American species and seven Asian species (He *et al.*, 2006). The triterpene glycoside cimigenol-3-*O*-arabinoside, cimifugin, and cimifugin-3-*O*-glucoside were suitable species-specific markers for the distinction of CR from the other species of *Cimicifuga* L. ex Wernisch. A rapid and reliable high-performance thin-layer chromatographic (HPTLC) method was developed for the identification of CR and the detection of its most common contaminants by fingerprint profiles (Ankli, Reich, and Steiner, 2008). The HPLC and LC-MS fingerprints for polyphenols and triterpene glycosides revealed the distinct patterns that made CR clearly distinguishable from most other species of *Actaea* L. (Jiang *et al.*, 2011). Cimifugin and cimracemoside F were found to be important to distinguish CR from most Asian species of *Actaea* L. CR products continue to be one of the most popular botanical supplements in the US and world markets, and the correct identification for the different species of *Actaea* L. is a key step for the good manufacturing practice.

Fifteen chemical markers, including 3 cimifugin derivatives, 11 triterpene glycosides and 1 alkaloid, were identified with HPLC-TOF-ESI-MS technique and principal component analysis (PCA), and the 16 species of *Actaea* L. were divided into three groups, Asian (seven species), North American (eight species) and *A. racemosa* groups (Ma *et al.*, 2011a). The adulteration of botanical supplements is a major problem that efficacy of the original product could be diminished and even toxicity could arise. Therefore, identification of marker compounds may help in the quality control and standardization of botanical supplements. Ultra performance liquid chromatography (UPLC) is a relatively new technique giving new possibilities in liquid chromatography, which is faster and needs less solvent than HPLC. UPLC-UV/ELSD and UPLC-MS were successfully used to analyze the

different CR market products as well as to distinguish other two species of *Actaea* L. (Avula *et al.*, 2009). Thin-layer chromatography (TLC) and combined TLC-bioluminescence (Bioluminex) are efficient, economical, and effective techniques which provided characteristic patterns and toxicity profiles for CR, *A. pachypoda* Ell. (white cohosh), *A. podocarpa* DC. (yellow cohosh), and other congeners (Verbitski *et al.*, 2008). In addition to identification, the fingerprint method provided insight into chemical interconversion processes occurring between the diverse triterpene glycosides contained in Cimicifugeae.

Molecular phylogeny and genomics

Sequence analysis of nuclear internal transcribed spacer (ITS) and fluorescence melting curve analysis of LightCycler real-time polymerase chain reaction (PCR) products were used to authenticate *C. foetida* from four substitutes: *C. heracleifolia*, *C. dahurica*, *C. acerina*, and *C. simplex* Wormsk (Xue, Li, and Wang, 2009). This method was expensive and did not resolve the phylogenetic relationship within Cimicifugeae. We retrieved the ITS and chloroplast (cp) DNA sequences from NCBI GenBank and constructed the phylogenetic tree. On the ITS tree (Fig. 2), *Beesia* Balf. f. et W. W. Sm. and *Anemonopsis* Siebold et Zucc. are basal to the intermingled *Actaea* L., *Cimicifuga* L. ex Wernisch., and *Souliea* Franch. sequences. There are two major clades. One clade includes *C. foetida*, *C. yunnanensis* Hsiao, *C. brachycarpa* Hsiao, *C. kashmiriana* J. Compton et Hedd., and *C. europaea* Schipcz., which is supported by the cp *trnL-F* tree; In the other clade, *C. heracleifolia* and *C. dahurica* are basal to other sequences, followed sequentially by *Souliea* Franch. and *C. nanchuanensis* Hsiao, an endemic endangered species in Sichuan, China. *C. acerina*, used as a folk medicine, grouped with two Japanese species, which is supported by the cp *matK* tree (not shown), while different *C. simplex* sequences cluster with distinct group. On the *trnL-F* tree, the group containing *C. foetida* and *C. yunnanensis* is closer to *C. heracleifolia* and *C. dahurica*, whereas *C. simplex* is closer to these Asian species. *C. yunnanensis* clusters with *C. foetida* (alternative names *C. mairei* and *C. frigida*) on both ITS and cp trees, implying that it might not be an independent species. *A. erythrocarpa* (Fisch.) Kom.

and *A. asiatica* Hara of China are close to *A. rubra* (Ait.) Willd. and *A. spicata* L. of North America respectively, and *A. racemosa* (black cohosh) is closer to these species than to other taxa. The reciprocal position of these American species and *Shengma* source plants is inverted on the ITS and cp trees. The phylogenetic position of *Souliea* Franch. varies greatly. These observations implied the extensive hybridization of the ancestral taxa during Cimicifugeae evolution.

A. racemosa cDNA libraries were constructed from young leaf, rhizome, and root tissues (Spiering *et al.*, 2011). Expressed sequence tags (ESTs) sequencing was performed and 1590 unigenes were assembled. Seventy putative secondary metabolism genes were identified, including 2, 3 oxidosqualene cyclase, BAHD-type acyltransferase, and tryptophan decarboxylase. This preliminary transcriptome study provided the initial insight into gene content and diversity in black cohosh, and generated the tools and resources for the detailed investigations of secondary metabolite genes and enzymes in this important medicinal plant. Seven microsatellite markers were found for *A. racemosa* (Pate *et al.*, 2012). Most of the loci cross-amplified in *A. pachypoda* Elliot, *A. podocarpa* DC., and *A. rubra*, indicating the utility of these markers for the genus. In the future, more microsatellite markers could be mined from the transcriptome and genomic survey datasets (Hao *et al.*, 2011; 2012), which will provide the tools for population genetic studies and molecular breeding.

The evolutionary history was inferred using the Neighbor-Joining method. The optimal tree with the sum of branch length = 0.512 682 92 was shown. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (500 replicates) was shown next to the branches. The tree was drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were computed using the Maximum Composite Likelihood method and were in the units of the number of base substitutions per site. The rate variation among sites was modeled with a γ distribution (shape parameter = 6.18). The differences in the composition bias among sequences were considered in evolutionary comparisons.

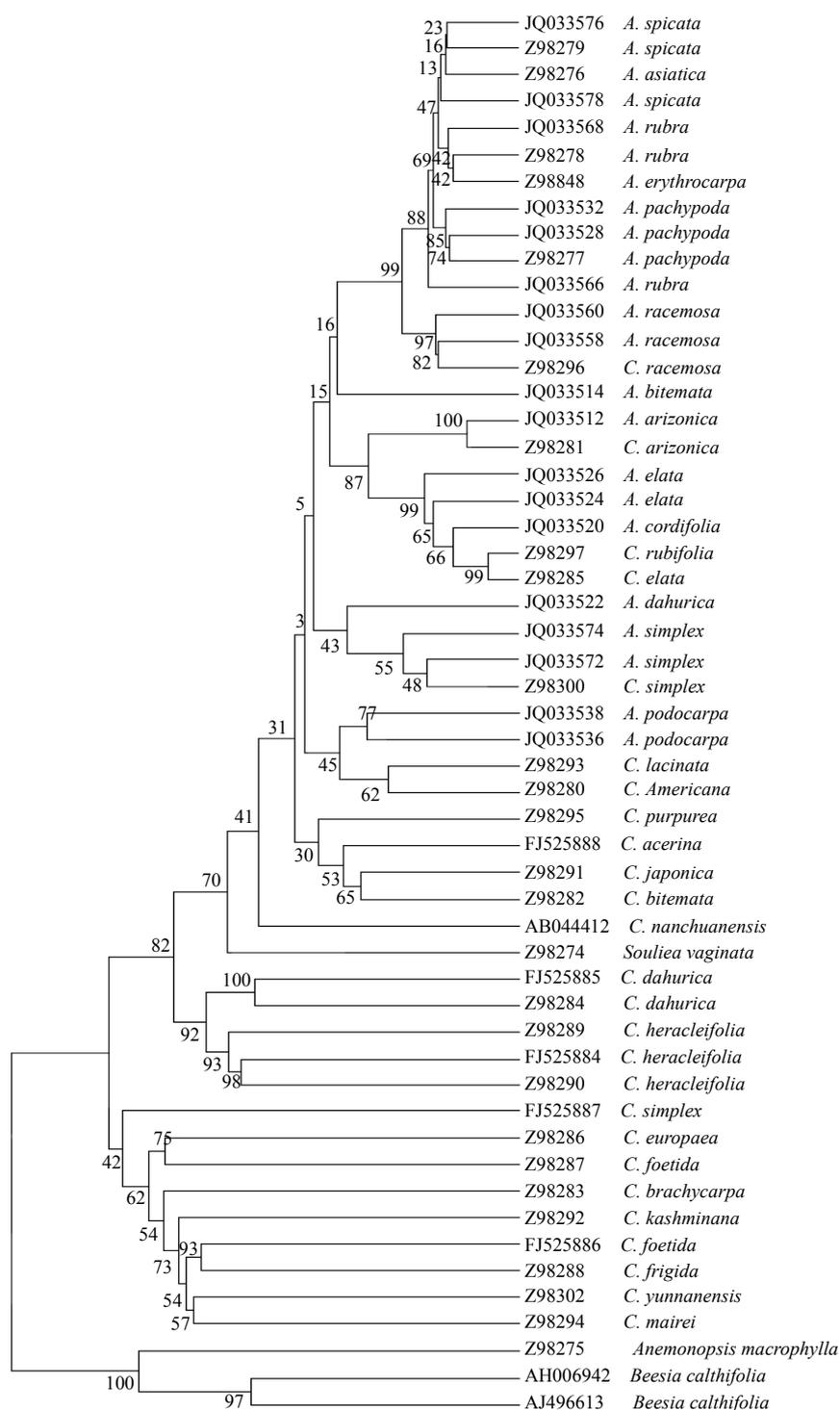


Fig. 2 Evolutionary relationships of Cimicifugeae ITS sequences

The analysis involved 53 nucleotide sequences. All ambiguous positions were removed for each sequence pair. There were a total of 754 positions in the final dataset. Evolutionary analyses were conducted in MEGA5 (Tamura *et al*, 2011). *C. foetida*, *C. dahurica*, and *C. heracleifolia* are officially listed in *China Pharmacopoeia* as “Shengma”.

Conclusion

The significant advances in the studies on pharmaceutical components, biological activities, taxonomy, and phylogeny of Cimicifugeae plants are summarized in this review. Recently more triterpenoid saponins have been found in Cimicifugeae plants, but their biosynthetic pathway has not been explored, which

hinders the cell-based production of useful secondary metabolites. Other secondary metabolites of Cimicifugeae plants are less studied and thus might be intriguing the topics in the future. Metabolomic and genomic studies of Cimicifugeae plants are just in their infancy. Various omics techniques should be put into full play in the drug research and development of Cimicifugeae.

References

- Ahn BS, Yang M, Jang H, Lee HJ, Moon C, Kim JC, Jung U, Jo SK, Jang JS, Kim SH, 2012. Evaluation of the antiosteoporotic potential of *Cimicifuga heracleifolia* in female mice. *Phytother Res* 26(5): 663-668.
- Ali Z, Khan SI, Fronczek FR, Khan IA, 2007a. 9,10-Seco-9,19-cyclolanostane arabinosides from the roots of *Actaea podocarpa*. *Phytochemistry* 68(3): 373-382.
- Ali Z, Khan SI, Khan IA, 2007. New cycloartane-type triterpene arabinosides from the roots of *Actaea podocarpa* and their biological study. *Planta Med* 73(7): 699-703.
- Ali Z, Khan SI, Pawar RS, Ferreira D, Khan IA, 2007b. 9,19-cyclolanostane derivatives from the roots of *Actaea pachypoda*. *J Nat Prod* 70(1): 107-110.
- Alves DL, Lima SM, da Silva CR, Galvão MA, Shanaider A, de Almeida Prado RA, Aoki T, 2008. Effects of *Trifolium pratense* and *Cimicifuga racemosa* on the endometrium of Wistar rats. *Maturitas* 61(4): 364-370.
- Ankli A, Reich E, Steiner M, 2008. Rapid high-performance thin-layer chromatographic method for detection of 5% adulteration of black cohosh with *Cimicifuga foetida*, *C. heracleifolia*, *C. dahurica*, or *C. americana*. *J AOAC Int* 91(6): 1257-1264.
- Avula B, Wang YH, Smillie TJ, Khan IA, 2009. Quantitative determination of triterpenoids and formononetin in rhizomes of black cohosh (*Actaea racemosa*) and dietary supplements by using UPLC-UV/ELS detection and identification by UPLC-MS. *Planta Med* 75(4): 381-386.
- Bai WP, Wang SY, Liu JL, Geng L, Hu LN, Zhang ZL, Chen SL, Zheng SR, 2009. Efficacy and safety of remifemin compared to tibolone for controlling of perimenopausal symptoms. *Chin J Obstet Gynecol* 44(8): 597-600.
- Campos LB, Gilgioni EH, Garcia RF, Brito Mdo N, Natali MR, Ishii-Iwamoto EL, Salgueiro-Pagadigorria CL, 2012. *Cimicifuga racemosa* impairs fatty acid β -oxidation and induces oxidative stress in livers of ovariectomized rats with renovascular hypertension. *Free Radic Biol Med* 53(4): 680-689.
- Choi EM, 2011. Deoxyactein isolated from *Cimicifuga racemosa* protects osteoblastic MC3T3-E1 cells against antimycin A-induced cytotoxicity. *J Appl Toxicol* doi: 10.1002/jat.1784
- Choi EM, 2013. Deoxyactein stimulates osteoblast function and inhibits bone-resorbing mediators in MC3T3-E1 cells. *J Appl Toxicol* 33(3): 190-195
- Cicek SS, Khom S, Taferner B, Hering S, Stuppner H, 2010. Bioactivity-guided isolation of GABA(A) receptor modulating constituents from the rhizomes of *Actaea racemosa*. *J Nat Prod* 73(12): 2024-2028.
- Compton JA, Culham A, 2002. Phylogeny and circumscription of Tribe Actaeae (Ranunculaceae). *Syst Bot* 27(3): 502-511.
- Dan C, Liang J, Zhou Y, Ding L, 2009. Cycloartane triterpenoid of *Cimicifuga foetida*. *Chin J Chin Mater Med* 34(15): 1930-1934.
- Dan C, Zhou Y, Ye D, Peng S, Ding L, Gross ML, Qiu SX, 2007. Cimicifugadine from *Cimicifuga foetida*, a new class of triterpene alkaloids with novel reactivity. *Org Lett* 9(9): 1813-1816.
- Daniels AL, van Slambrouck S, Lee RK, Arguello TS, Browning J, Pullin MJ, Kornienko A, Steelant WF, 2006. Effects of extracts from two native American plants on proliferation of human breast and colon cancer cell lines *in vitro*. *Oncol Rep* 15(5): 1327-1331.
- Davis VL, Jayo MJ, Ho A, Kotlarczyk MP, Hardy ML, Foster WG, Hughes CL, 2008. Black cohosh increases metastatic mammary cancer in transgenic mice expressing c-erbB2. *Cancer Res* 68(20): 8377-8383.
- Einbond LS, Soffritti M, Degli Esposti D, Tibaldi E, Lauriola M, Bua L, He K, Genovese G, Su T, Huggins L, Wang X, Roller M, Wu HA, 2012. Chemopreventive potential of black cohosh on breast cancer in Sprague-Dawley rats. *Anticancer Res* 32(1): 21-30.
- Einbond LS, Soffritti M, Esposti DD, Park T, Cruz E, Su T, Wu HA, Wang X, Zhang YJ, Ham J, Goldberg JJ, Kronenberg F, Vladimirova A, 2009. Actein activates stress- and statin-associated responses and is bioavailable in Sprague-Dawley rats. *Fundam Clin Pharmacol* 23(3): 311-321.
- Einbond LS, Su T, Wu HA, Friedman R, Wang X, Jiang B, Hagan T, Kennelly EJ, Kronenberg F, Weinstein IB, 2007. Gene expression analysis of the mechanisms whereby black cohosh inhibits human breast cancer cell growth. *Anticancer Res* 27(2): 697-712.
- Einbond LS, Ye WC, He K, Wu HA, Cruz E, Roller M, Kronenberg F, 2008. Growth inhibitory activity of extracts and compounds from *Cimicifuga* species on human breast cancer cells. *Phytomedicine* 15(6/7): 504-511.
- Fan YS, Yao Z, Zhang YW, Duan HQ, 2009. Six new cycloartane triterpene glycosides from *Actaea asiatica*. *J Asian Nat Prod Res* 11(7): 588-596.
- Gao JC, Peng Y, Yang MS, Xiao PG, 2008. A preliminary phylogenetic study of tribe Cimicifugeae (Ranunculaceae). *J Syst Evol* 46: 516-536.
- Gaube F, Wolf S, Pusch L, Kroll TC, Hamburger M, 2007. Gene expression profiling reveals effects of *Cimicifuga racemosa* (L.) NUTT. (black cohosh) on the estrogen receptor positive human breast cancer cell line MCF-7. *BMC Pharmacol* 7: 11.
- Gödecke T, Lankin DC, Nikolic D, Chen SN, van Breemen RB, Farnsworth NR, Pauli GF, 2009. Guanidine alkaloids and Pictet-Spengler adducts from black cohosh (*Cimicifuga racemosa*). *J Nat Prod* 72(3): 433-437.
- Gödecke T, Nikolic D, Lankin DC, Chen SN, Powell SL, Dietz B, Bolton JL, van Breemen RB, Farnsworth NR, Pauli GF, 2009. Phytochemistry of cimicifugic acids and associated bases in *Cimicifuga racemosa* root extracts. *Phytochem Anal* 20(2): 120-133.
- Guo LY, Joo EJ, Son KH, Jeon SJ, Jang S, Shin EM, Zhou HY, Kim YS, 2009. Cimicide E arrests cell cycle and induces cell apoptosis in gastric cancer cells. *Arch Pharm Res* 32(10): 1385-1392.
- Hao DC, Ma P, Mu J, Chen SL, Xiao PG, Peng Y, Huo L, Xu LJ, Sun C, 2012. De novo characterization of the root transcriptome of a

- traditional Chinese medicinal plant *Polygonum cuspidatum*. *Sci China Life Sci* 55(5): 452-466.
- Hao DC, Yang L, Xiao PG, 2011. The first insight into the *Taxus* genome via fosmid library construction and end sequencing. *Mol Genet Genomics* 285(3): 197-205.
- He K, Pauli GF, Zheng B, Wang H, Bai N, Peng T, Roller M, Zheng Q, 2006. *Cimicifuga* species identification by high performance liquid chromatography-photodiode array/mass spectrometric/evaporative light scattering detection for quality control of black cohosh products. *J Chromatogr A* 1112(1/2): 241-254.
- Ho SH, Singh M, Holloway AC, Crankshaw DJ, 2011. The effects of commercial preparations of herbal supplements commonly used by women on the biotransformation of fluorogenic substrates by human cytochromes P450. *Phytother Res* 25(7): 983-989.
- Huang Y, Jiang B, Nuntanakorn P, Kennelly EJ, Shord S, Lawal TO, Mahady GB, 2010. Fukinolic acid derivatives and triterpene glycosides from black cohosh inhibit CYP isozymes, but are not cytotoxic to Hep-G2 cells *in vitro*. *Curr Drug Saf* 5(2): 118-124.
- Huh JE, Shin YJ, Baek YH, Lee JD, Choi DY, Park DS, 2011. Combined prescription (OAH19T) of *Aralia cordata* Thunb and *Cimicifuga heracleifolia* Komar and its major compounds inhibit matrix proteinases and vascular endothelial growth factor through the regulation of mitogen-activated protein kinase pathway. *J Ethnopharmacol* 135(2): 414-421.
- Iwanaga A, Kusano G, Warashina T, Miyase T, 2010a. Hyaluronidase inhibitors from “*Cimicifugae Rhizoma*” (a mixture of the rhizomes of *Cimicifuga dahurica* and *C. heracleifolia*). *J Nat Prod* 73(4): 573-578.
- Iwanaga A, Kusano G, Warashina T, Miyase T, 2010b. Phenolic constituents of the aerial parts of *Cimicifuga simplex* and *Cimicifuga japonica*. *J Nat Prod* 73(4): 609-612.
- Jamrůz MK, Jamrůz MH, Dobrowolski JC, Gliński JA, Gleńsk MH, 2012. One new and six known triterpene xylosides from *Cimicifuga racemosa*: FT-IR, Raman and NMR studies and DFT calculations. *Spectrochim Acta A Mol Biomol Spectrosc* 93: 10-18.
- Jamrůz MK, Jamrůz MH, Dobrowolski JC, Gliński JA, Davey MH, Wawer I, 2011. Novel and unusual triterpene from Black Cohosh. Determination of structure of 9,10-seco-9,19-cyclolanostane xyloside (cimipodocarpaside) by NMR, IR and Raman spectroscopy and DFT calculations. *Spectrochim Acta A Mol Biomol Spectrosc* 78(1): 107-112.
- Jang JY, Lee JH, Kang BW, Chung KT, Choi YH, Choi BT, 2009. Dichloromethane fraction of *Cimicifuga heracleifolia* decreases the level of melanin synthesis by activating the ERK or AKT signaling pathway in B16F10 cells. *Exp Dermatol* 18(3): 232-237.
- Jarry H, Stromeier S, Wuttke W, Nahrstedt A, 2007. Petasiphenone, a phenol isolated from *Cimicifuga racemosa*, *in vitro* inhibits proliferation of the human prostate cancer cell line LNCaP. *Planta Med* 73(2): 184-187.
- Jiang B, Kronenberg F, Nuntanakorn P, Qiu MH, Kennelly EJ, 2006. Evaluation of the botanical authenticity and phytochemical profile of black cohosh products by high-performance liquid chromatography with selected ion monitoring liquid chromatography-mass spectrometry. *J Agric Food Chem* 54(9): 3242-3253.
- Jiang B, Ma C, Motley T, Kronenberg F, Kennelly EJ, 2011. Phytochemical fingerprinting to thwart black cohosh adulteration: A 15 *Actaea* species analysis. *Phytochem Anal* 22(4): 339-351.
- Kaminski CN, Ferrey SL, Lowrey T, Guerra L, VAN Slambrouck S, Steelant WF, 2010. *In vitro* anticancer activity of *Anemopsis californica*. *Oncol Lett* 1(4): 711-715.
- Kapur P, Wuttke W, Seidlova-Wuttke D, 2010. The *Cimicifuga racemosa* special extract BNO 1055 prevents hot flashes in ovariectomized rats. *Phytomedicine* 17(11): 890-894.
- Kim EY, Lee YJ, Rhyu MR, 2011. Black cohosh (*Cimicifuga racemosa*) relaxes the isolated rat thoracic aorta through endothelium-dependent and -independent mechanisms. *J Ethnopharmacol* 138(2): 537-542.
- Kolios L, Schumann J, Sehmisch S, Rack T, Tezval M, Seidlova-Wuttke D, Frosch KH, Stuermer KM, Stuermer EK, 2010. Effects of black cohosh (*Cimicifuga racemosa*) and estrogen on metaphyseal fracture healing in the early stage of osteoporosis in ovariectomized rats. *Planta Med* 76(9): 850-857.
- Kuang H, Su Y, Wang Q, Wu L, Yang B, Wang Z, Xia Y, 2012. Three new cycloartenol glycosides from the roots of *Cimicifuga simplex*. *Planta Med* 78(6): 622-625.
- Kuang H, Su Y, Yang B, Xia Y, Wang Q, Wang Z, Yu Z, 2011. Three new cycloartenol triterpenoid saponins from the roots of *Cimicifuga simplex* Wormsk. *Molecules* 16(6): 4348-4357.
- Laakmann E, Grajecki D, Doege K, Zu Eulenburg C, Buhling KJ, 2012. Efficacy of *Cimicifuga racemosa*, *Hypericum perforatum* and *Agnus castus* in the treatment of climacteric complaints: A systematic review. *Gynecol Endocrinol* 28(9): 703-709.
- Lee JH, Cuong TD, Kwack SJ, Seok JH, Lee JK, Jeong JY, Woo MH, Choi JS, Lee HK, Min BS, 2012. Cycloartane-type triterpene glycosides from the rhizomes of *Cimicifuga heracleifolia* and their anticomplementary activity. *Planta Med* 78(12): 1391-1394.
- Li J, Gödecke T, Chen SN, Imai A, Lankin DC, Farnsworth NR, Pauli GF, van Breemen RB, Nikolić D, 2011. *In vitro* metabolic interactions between black cohosh (*Cimicifuga racemosa*) and tamoxifen via inhibition of cytochromes P450 2D6 and 3A4. *Xenobiotica* Aug9: [Epub ahead of print]
- Li JX, Liu J, He CC, Yu ZY, Du Y, Kadota S, Seto H, 2007. Triterpenoids from *Cimicifugae rhizoma*, a novel class of inhibitors on bone resorption and ovariectomy-induced bone loss. *Maturitas* 58(1): 59-69.
- Lu B, Fan YS, Duan HQ, 2008. A new cycloartane triterpene saponin from rhizome of *Actaea asiatica*. *China J Chin Mater Med* 33(13): 1558-1561.
- Lu L, Chen J, Nian Y, Sun Y, Qiu M, 2009. Trinor-cycloartane glycosides from the rhizomes of *Cimicifuga foetida*. *Molecules* 14(4): 1578-1584.
- Lu L, Chen JC, Li Y, Qing C, Wang YY, Nian Y, Qiu MH, 2012. Studies on the constituents of *Cimicifuga foetida* collected in Guizhou Province and their cytotoxic activities. *Chem Pharm Bull* 60(5): 571-577.
- Lu L, Chen JC, Song HJ, Li Y, Nian Y, Qiu MH, 2010. Five new triterpene bisglycosides with acyclic side chains from the rhizomes of *Cimicifuga foetida* L. *Chem Pharm Bull* 58(5): 729-733.
- Ma C, Kavalier AR, Jiang B, Kennelly EJ, 2011a. Metabolic profiling of *Actaea* species extracts using high performance liquid chromatography coupled with electrospray ionization time-of-flight mass spectrometry. *J Chromatogr A* 1218(11): 1461-1476.

- Ma X, Zhang H, Wang K, Yang L, Qin L, Bai W, Guan Y, Jia J, Kang J, Zhou C, 2011b. Effects of an isopropanolic-aqueous black cohosh extract on central body temperature of ovariectomized rats. *J Ethnopharmacol* 138(1): 156-161.
- Medina AL, Lucero ME, Holguin FO, Estell RE, Posakony JJ, Simon J, O'Connell MA, 2005. Composition and antimicrobial activity of *Anemopsis californica* leaf oil. *J Agric Food Chem* 53(22): 8694-8698.
- Medina-Holguín AL, Holguín FO, Micheletto S, Goehle S, Simon JA, O'Connell MA, 2008. Chemotypic variation of essential oils in the medicinal plant, *Anemopsis californica*. *Phytochemistry* 69(4): 919-927.
- Mercado-Feliciano M, Cora MC, Witt KL, Granville CA, Hejtmancik MR, Fomby L, Knostman KA, Ryan MJ, Newbold R, Smith C, Foster PM, Vallant MK, Stout MD, 2012. An ethanolic extract of black cohosh causes hematological changes but not estrogenic effects in female rodents. *Toxicol Appl Pharmacol* 263(2): 138-147.
- Moon L, Ha YM, Jang HJ, Kim HS, Jun MS, Kim YM, Lee YS, Lee DH, Son KH, Kim HJ, Seo HG, Lee JH, Kim YS, Chang KC, 2011. Isoimperatorin, cimicidine E and 23-O-acetylshengmanol-3-xyloside from *Cimicifugae Rhizome* inhibit TNF- α -induced VCAM-1 expression in human endothelial cells: involvement of PPAR- γ upregulation and PI3K, ERK1/2, and PKC signal pathways. *J Ethnopharmacol* 133(2): 336-344.
- Mun L, Jun MS, Kim YM, Lee YS, Kim HJ, Seo HG, Lee JH, Son KH, Lee DH, Kim YS, Park K, Chang KC, 2011. 7,8-didehydrocimigenol from *Cimicifugae Rhizoma* inhibits TNF- α -induced VCAM-1 but not ICAM-1-expression through upregulation of PPAR- γ in human endothelial cells. *Food Chem Toxicol* 49(1): 166-172.
- Nadaoka I, Watanabe K, Yasue M, Sami M, Kitagawa Y, Mimaki Y, 2012c. Oral administration of *Cimicifuga racemosa* extract attenuates immobilization stress-induced reactions. *Nat Prod Commun* 7(1): 15-18.
- Nadaoka I, Yasue M, Kitagawa Y, Koga Y, 2012a. Oral administration of *Cimicifuga racemosa* extract attenuates psychological and physiological stress responses. *Biomed Res* 33(3): 145-152.
- Nadaoka I, Yasue M, Sami M, Kitagawa Y, 2012b. Oral administration of *Cimicifuga racemosa* extract affects immobilization stress-induced changes in murine cerebral monoamine metabolism. *Biomed Res* 33(2):133-137.
- Nian Y, Wang HY, Su J, Zhou L, Qiu MH, 2012. A cytotoxic 4 α -methyl steroid from the aerial parts of *Cimicifuga foetida* L. *Fitoterapia* 83(2): 293-297.
- Nian Y, Zhang XM, Li Y, Wang YY, Chen JC, Lu L, Zhou L, Qiu MH, 2011. Cycloartane triterpenoids from the aerial parts of *Cimicifuga foetida* Linnaeus. *Phytochemistry* 72(11/12): 1473-1481.
- Nian Y, Zhang YL, Chen JC, Lu L, Qiu MH, Qing C, 2010. Cytotoxic chemical constituents from the roots of *Cimicifuga foetida*. *J Nat Prod* 73(2): 93-98.
- Nishida M, Yoshimitsu H, 2011. Six new cycloartane glycosides from cimicifuga rhizome. *Chem Pharm Bull* 59(10): 1243-1249.
- Nuntanakorn P, Jiang B, Einbond LS, Yang H, Kronenberg F, Weinstein IB, Kennelly EJ, 2006. Polyphenolic constituents of *Actaea racemosa*. *J Nat Prod* 69(3): 314-318.
- Nuntanakorn P, Jiang B, Yang H, Cervantes-Cervantes M, Kronenberg F, Kennelly EJ, 2007. Analysis of polyphenolic compounds and radical scavenging activity of four American *Actaea* species. *Phytochem Anal* 18(3): 219-228.
- Pan RL, Chen DH, Si JY, Zhao XH, 2007. Cimifoetisides VI and VII two new cyclolanostanol triterpene glycosides from the aerial parts of *Cimicifuga foetida*. *J Asian Nat Prod Res* 9(2): 97-102.
- Pan RL, Chen DH, Si JY, Zhao XH, Li Z, Cao L, 2009. Immunosuppressive effects of new cyclolanostane triterpene diglycosides from the aerial part of *Cimicifuga foetida*. *Arch Pharm Res* 32(2): 185-190.
- Pang X, Cheng J, Krausz KW, Guo DA, Gonzalez FJ, 2011. Pregnane X receptor-mediated induction of Cyp3a by black cohosh. *Xenobiotica* 41(2): 112-123.
- Pate SJ, Clement JA, McCoy JA, Lance SL, Mathews KG, 2012. Development and characterization of microsatellite markers for *Actaea racemosa* (black cohosh, Ranunculaceae). *Am J Bot* 99(7): e274-e276.
- Powell SL, Gödecke T, Nikolic D, Chen SN, Ahn S, Dietz B, Farnsworth NR, van Breemen RB, Lankin DC, Pauli GF, Bolton JL, 2008. *In vitro* serotonergic activity of black cohosh and identification of N(omega)-methylserotonin as a potential active constituent. *J Agric Food Chem* 56(24): 11718-11726.
- Qiu M, Kim JH, Lee HK, Min BS, 2006. Anticomplement activity of cycloartane glycosides from the rhizome of *Cimicifuga foetida*. *Phytother Res* 20(11): 945-948.
- Rachón D, Vortherms T, Seidlová-Wuttke D, Wuttke W, 2008. Effects of black cohosh extract on body weight gain, intra-abdominal fat accumulation, plasma lipids and glucose tolerance in ovariectomized Sprague-Dawley rats. *Maturitas* 60(3/4): 209-215.
- Ross SM, 2012. Menopause: A standardized isopropanolic black cohosh extract (remifemin) is found to be safe and effective for menopausal symptoms. *Holist Nurs Pract* 26(1): 58-61.
- Rostock M, Fischer J, Mumm A, Stammwitz U, Saller R, Bartsch HH, 2011. Black cohosh (*Cimicifuga racemosa*) in tamoxifen-treated breast cancer patients with climacteric complaints - a prospective observational study. *Gynecol Endocrinol* 27(10): 844-848.
- Schmid D, Woehs F, Svoboda M, Thalhammer T, Chiba P, Moeslinger T, 2009. Aqueous extracts of *Cimicifuga racemosa* and phenol-carboxylic constituents inhibit production of proinflammatory cytokines in LPS-stimulated human whole blood. *Can J Physiol Pharmacol* 87(11): 963-972.
- Seidlova-Wuttke D, Eder N, Stahnke V, Kammann M, Stecher G, Haunschild J, Wessels JT, Wuttke W, 2012a. *Cimicifuga racemosa* and its triterpene-saponins prevent the metabolic syndrome and deterioration of cartilage in the knee joint of ovariectomized rats by similar mechanisms. *Phytomedicine* 19(8/9): 846-853.
- Seidlova-Wuttke D, Jarry H, Wuttke W, 2009. Effects of estradiol benzoate, raloxifen and an ethanolic extract of *Cimicifuga racemosa* in nonclassical estrogen regulated organs of ovariectomized rats. *Planta Med* 75(12): 1279-1285.
- Seidlova-Wuttke D, Stecher G, Kammann M, Haunschild J, Eder N, Stahnke V, Wessels J, Wuttke W, 2012b. Osteoprotective effects of *Cimicifuga racemosa* and its triterpene-saponins are responsible for reduction of bone marrow fat. *Phytomedicine* 19(10): 855-860.

- Seidlová-Wuttke D, Thelen P, Wuttke W, 2006. Inhibitory effects of a black cohosh (*Cimicifuga racemosa*) extract on prostate cancer. *Planta Med* 72(6): 521-526.
- Shahin AY, Ismail AM, Zahran KM, Makhoulf AM, 2008. Adding phytoestrogens to clomiphene induction in unexplained infertility patients—a randomized trial. *Reprod Biomed Online* 16(4): 580-588.
- Shen JW, 2010. *Studies on the Chemical Constituents of Two Traditional Medicine*. Chinese Academy of Sciences, Northwest Institute of Plateau Biology: Xining.
- Soler MC, Molina JL, Díaz HA, Pinto VC, Barrios YL, He K, Roller M, Weinstein-Oppenheimer CR, 2011. Effect of the standardized *Cimicifuga foetida* extract on Hsp 27 expression in the MCF-7 cell line. *Biol Res* 44(3): 243-249.
- Spiering MJ, Urban LA, Nuss DL, Gopalan V, Stoltzfus A, Eisenstein E, 2011. Gene identification in black cohosh (*Actaea racemosa* L.): Expressed sequence tag profiling and genetic screening yields candidate genes for production of bioactive secondary metabolites. *Plant Cell Rep* 30(4): 613-629.
- Stute P, Nisslein T, Götte M, Kamischke A, Kiesel L, Klockenbusch W, 2007. Effects of black cohosh on estrogen biosynthesis in normal breast tissue *in vitro*. *Maturitas* 57(4): 382-391.
- Sun LR, Qing C, Zhang YL, Jia SY, Li ZR, Pei SJ, Qiu MH, Gross ML, Qiu SX, 2007. Cimicifoetisides A and B, two cytotoxic cycloartane triterpenoid glycosides from the rhizomes of *Cimicifuga foetida*, inhibit proliferation of cancer cells. *Beilstein J Org Chem* 3: 3.
- Sun LR, Yan J, Zhou L, Li ZR, Qiu MH, 2011. Two new triterpene glycosides with monomethyl malonate groups from the rhizome of *Cimifuga foetida* L. *Molecules* 16(7): 5701-5708.
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S, 2011. MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol Biol Evol* 28: 2731-2739.
- Teschke R, Schwarzenboeck A, Schmidt-Taenzer W, Wolff A, Hennermann KH, 2011. Herb induced liver injury presumably caused by black cohosh: A survey of initially purported cases and herbal quality specifications. *Ann Hepatol* 10(3): 249-259.
- Tian Z, Pan R, Chang Q, Si J, Xiao PG, Wu E, 2007a. *Cimicifuga foetida* extract inhibits proliferation of hepatocellular cells via induction of cell cycle arrest and apoptosis. *J Ethnopharmacol* 114(2): 227-233.
- Tian Z, Si JY, Chang Q, Zhou L, Chen S, Xiao PG, Wu E, 2007b. Antitumor activity and mechanisms of action of total glycosides from aerial part of *Cimicifuga dahurica* targeted against hepatoma. *BMC Cancer* 7: 237.
- Tian Z, Zhou L, Huang F, Chen S, Yang J, Wu E, Xiao PG, Yang M, 2006. Anti-cancer activity and mechanisms of 25-anhydrocimigenol-3-O-beta-D-xylopyranoside isolated from *Souliea vaginata* on hepatomas. *Anticancer Drugs* 17(5): 545-551.
- Verbitski SM, Gourdin GT, Ikenouye LM, McChesney JD, Hildreth J, 2008. Detection of *Actaea racemosa* adulteration by thin-layer chromatography and combined thin-layer chromatography-bioluminescence. *JAOAC Int* 91(2): 268-275.
- Wang KC, Chang JS, Chiang LC, Lin CC, 2012a. *Cimicifuga foetida* L. inhibited human respiratory syncytial virus in HEP-2 and A549 cell lines. *Am J Chin Med* 40(1): 151-162.
- Wang KC, Chang JS, Lin LT, Chiang LC, Lin CC, 2012b. Antiviral effect of cimicifugin from *Cimicifuga foetida* against human respiratory syncytial virus. *Am J Chin Med* 40(5): 1033-1045.
- Wang W, Lu AM, Ren Y, Endress ME, Chen ZD, 2009. Phylogeny and classification of Ranunculales: Evidence from four molecular loci and morphological data. *Persp Plant Ecol Evol Syst* 11(2): 81-110.
- Wang X, Li X, Chen D, 2011. Evaluation of antioxidant activity of isoferulic acid *in vitro*. *Nat Prod Commun* 6(9): 1285-1288.
- Xue CY, Li DZ, Wang QZ, 2009. Application of LightCycler polymerase chain reaction and melting curve analysis to the authentication of the traditional Chinese medicinal plant *Cimicifuga foetida*. *Planta Med* 75(8): 873-875.
- Yang CL, Chik SC, Li JC, Cheung BK, Lau AS, 2009. Identification of the bioactive constituent and its mechanisms of action in mediating the anti-inflammatory effects of black cohosh and related *Cimicifuga* species on human primary blood macrophages. *J Med Chem* 52(21): 6707-6715.
- Yawata A, Matsuhashi Y, Kato H, Uemura K, Kusano G, Ito J, Chikuma T, Hojo H, 2009. Inhibition of nucleoside transport and synergistic potentiation of methotrexate cytotoxicity by cimicifugoside, a triterpenoid from *Cimicifuga simplex*. *Eur J Pharm Sci* 38(4): 355-361.
- Yoshimitsu H, Nishida M, Nohara T, 2007. Three new 15,16-seco-cycloartane glycosides from *Cimicifuga Rhizome*. *Chem Pharm Bull* 55(5): 789-792.
- Zadoyan G, Fuhr U, 2012. Phenotyping studies to assess the effects of phytopharmaceuticals on *in vivo* activity of main human cytochrome p450 enzymes. *Planta Med* 78(13): 1428-1457.
- Zhang H, Ma XY, Yang MK, Wang K, Yang LY, Zhu SN, Jia J, Qin LH, Bai WP, 2012. Effects of black cohosh and estrogen on the hypothalamic nuclei of ovariectomized rats at different temperatures. *J Ethnopharmacol* 142(3): 769-775.