

New Collection of Crude Drugs in *Chinese Pharmacopoeia 2010* II. *Sankezhen* (*Berberis* spp.)

DAN Yang¹, LIU Yan-ze¹, PENG Yong¹, QIAN Zhong-zhi², XIAO Pei-gen^{1*}

1. Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences, Beijing 100193, China

2. Chinese Pharmacopoeia Commission, Beijing 100061, China

Abstract: *Sankezhen* (*Berberidis Radix*) is a traditional Chinese materia medica, cold in nature and bitter in taste, for treating syndromes of liver, stomach, and large intestinal meridians, in which berberine and berbamine are the major pharmacological components. *Sankezhen* has been readmitted in *Chinese Pharmacopoeia 2010* following the 1977 version as the roots of *Berberis* spp. e.g. *B. soulieana*, *B. wilsonae*, *B. poiretii*, *B. verna*, etc. Recent studies showed that *Berberis* spp. were potential phytomedicines with multiple spectrums therapeutic effects and various pharmaceutical parts. Here we reviewed *Sankezhen* in traditional use and phytochemistry, and its major active components berberine and berbamine with potential bioactivities recently discovered, such as antitumor, antidiabetic, antihyperlipidemic, anti-arrhythmic, and neuro-protective activities. It is necessary to mature the quality assessment of *Sankezhen* as a new admission of *Chinese Pharmacopoeia 2010*. Other parts of *Berberis* spp. should be investigated to better develop this herb in medicinal usage.

Key words: alkaloids; *Berberidis Radix*; berberine; berbamine; *Chinese Pharmacopoeia*; ethnopharmacology

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Berberis L. includes about 500 species of spiny or unarmed, evergreen or deciduous shrubs native to Asia, Europe, North Africa, and to North, Central, and South America. Over 200 species spread in the Southwest and Northwest of China, many of which are called *Sankezhen* (three spines) in folk Chinese due to the characteristics of three spines fasciculate on stems. Xiao *et al* (1974) reported that there are only 40–50 medicinal species in this genus which are abundant and widely distributed in China.

Sankezhen (*Berberidis Radix*) has been readmitted in *Chinese Pharmacopoeia 2010* (Pharmacopoeia Committee of P. R. China, 2010) following the 1977 version (Pharmacopoeia Committee of P. R. China, 1977) due to its multiple spectrum therapeutic effects and wide distribution in China. The botanical origins are authorized as dried roots of *Berberis* spp. e.g. *B. soulieana* Schneid., *B. wilsonae* Hemsl., *B. poiretii* Schneid., and *B. verna* Schneid., etc, which is yet to clarify the botanical origins to date. So, the so-called *Sankezhen* as medicine here and thereafter means the

radix of many species of *Berberis* spp. which could be used as the same purpose. Fortunately, the four species in *Chinese Pharmacopoeia 2010* might be potential candidates due to their abundance as the representatives in various regions of China such as Southwest, Northwest, and North China. For example, *B. soulieana* is largely spread in Qinling and Daba mountains, and in Sichuan, Hubei, Gansu, and Shanxi Provinces. *B. wilsonae* is abundant in Southwest China including Sichuan, Yunnan, Guizhou, Gansu, Hubei Provinces, and Tibet. *B. poiretii* grows in North China, such as forests of Daxing'anling and Xiaoxing'anling, and Hebei, Shandong, Shanxi, Shanxi Provinces, and Inner Mongolia, etc. *B. verna* is popular in Northwest China, especially in Gansu and Qinghai Provinces and Xinjiang Autonomous Region. However, few studies have been intensively carried out on their phytochemistry.

Since *Berberis* spp. spread worldwide and are used for medicinal purpose, here we reviewed *Sankezhen* in traditional use, phytochemistry, and pharmacology recently discovered. Since *Sankezhen* is one of the new

* Corresponding author: Xiao PG

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admissions of *Chinese Pharmacopeia 2010*, it is in great needs to improve current quality assessment upon this herb. In addition, we briefly introduced other bioactive parts of *Berberis* spp. and the responsible compounds in order to better develop *Berberis* spp. in medicinal use.

Botanical description

Berberis L. is described in *Flora of China* as follows: shrubs, evergreen or deciduous. Branches glabrous or tomentose, spinose or not; spines simple or usually 3–5-fid. Leaves on short shoots, simple, alternate, usually petiolate. Inflorescences solitary or fascicled flowers, racemes, umbels, or panicles. Flowers 3-merous; bracteoles usually 3, caducous, scalelike. Sepals 6, rarely 3 or 9, yellow. Petals 6, yellow, bases nectariferous. Stamens opposite petals; anthers dehiscing by valves; pollen grains subspheroidal, exine reticulate. Ovary symmetrically club-shaped; ovules 1–12, rarely to 15, subbasal; styles very short. Fruit a berry, usually red, dark red, or black, globose, ellipsoid, oblong, ovoid, or obovoid. Seeds 1–10, tan to red-brown or black; aril absent. $2n = 14$ (Ying and Ying,).

Traditional use

Sankezhen has long been used for treating liver, stomach, and large intestinal meridian syndromes in traditional Chinese medicine (TCM) (Committee of *Chinese Herbacology*, 2004). It was first documented in *Tangbencao* that the plants were as small as pomegranate trees in height with yellow barks and bitter taste. *Bencao Shiyi* said that *Sankezhen* were alternative to *Phellodendri Chinensis Cortex* and *Coptidis Rhizoma* in TCM. They are similar in the following aspects, cold in nature and bitter in taste, clearing heat and drying dampness, purging fire for removing toxin, and strengthening the stomach, orally used in treating dysentery, gastroenteritis, hot eyes, toothache, aptha, sore throat, acute hepatitis, chronic cholecystitis, jaundice, and dyspepsia, and externally used to treat swelling and ulcer on the body, hot eyes, burn, eczema, and other skin infection diseases (Xiao *et al.*, 1974). However, *Sankezhen* is absent from traditional Chinese prescriptions. Roots and barks of the species in *Berberis* L. are also used as traditional

medicines in Korea, India, and Pakistan. *B. koreana* Palibin has been used to treat gastroenteritis, sore throats, fever, and conjunctivitis in Korea (Kim, Choi, and Lee, 2010). *B. asiatica* (Daruharidra) is an important ingredient of the Ayurvedic system in India (Andola *et al.*, 2010). The extracts from roots, leaves, and fruits of *B. lyceum* have been used against liver cirrhosis, hepatitis, colic disorder, chest cold, suppressed cough, and burned wound in Pakistan (Gulfraz *et al.*, 2008).

It is generally believed that alkaloids, principally berberine are the major compounds in *Sankezhen*. Berberine is a botanic drug in clinic to treat cholera, severe diarrhea, amoebiasis, and intestinal infections, which has been admitted as *Huangliansu* in *Chinese Pharmacopeia*. In recent decades, it has been demonstrated to possess antimicrobial and anticancer bioactivities, and it is also used for cardiovascular disease and central nervous disorder (Tang *et al.*, 2009). Although total synthesis of berberine was achieved in 1969, it is largely extracted from the plants of *Berberis* L., which are major natural resources of berberine upon the demand of pharmaceutical industry. In China, Xiao *et al.* (1974) reported that the good botanical resources for producing berberine were *B. gagnepainii* var. *lanceifolia*, *B. sanguine*, *B. soulieana*, *B. taronensis*, and *B. reticulate* due to higher yields of berberine (1.08%–3.04% of root dry weight) though the highest yields of berberine (3.95%–4.17 % of root dry weight) were found in *B. virgetorum*, *B. henryana*, and *B. polyantha*. The best botanical resources for the production of berberine rely on population biomass and berberine contents in plants.

Due to difference in chemical structures, berberine and berbamine are used differently in clinical practices. Berberine is commonly used in gastroenteritis, neonatal conjunctivitis, and tympanitis caused by various microbe with the single dose of 0.1–0.4 g by ig administration and as anti-arrhythmics with the single dose of 0.6–1 g, while berbamine, another major alkaloid in *Sankezhen*, is mainly used in stimulating normal hematopoiesis and anti-arrhythmics in clinical practice.

Chemical components in *Sankezhen* and other parts of *Berberis* spp.

The major compounds of *Sankezhen* are alkaloids,

e.g. berberine, berbamine, jatrorrhizin, palmatine (Fig. 1) and so on, which are also detected in shoots (e.g. trunks, barks, and stems). In addition, other classes of compounds have been isolated and identified in the rest parts of *Berberis* spp. (e.g. polyphenols, triterpens, biphenyls, saponin, pyrrole acids, and tannin) (Gulfraz *et al.*, 2008; Rashmia, Rajasekaranb, and Pant, 2008). Since the previous review (Rashmia, Rajasekaranb, and Pant, 2008) summed up the chemical constituents discovered before 2008, we show the alkaloids discovered in the latest four years (Table 1). The major components of roots, twigs, and barks are alkaloids containing aporphines (e.g. magnoflorine), proaporphines (e.g. pronuciferine and pronuciferine *N*-oxide) (Fajardo *et al.*, 2009), bisbenzylisoquinolines (e.g. tabienine, isotetrandrine, obamegine, and thaligrisine) (Suau *et al.*, 1998), pseudobenzylisoquinolines (e.g. dihydrorugosinone) (Valencia, Weiss, and Shamma, 1984), aporphinebenzylisoquinolines (e.g. 1-*O*-methylpakistanine, kalashine, chitraline, and pakistanine) (Hussain *et al.*, 1986), proaporphine-benzylisoquinolines (e.g. pakistannamine) (Hussain *et al.*, 1986), benzytetrahydroisoquinolines (e.g. reticuline, glaucine, and thalicmidine) (Karimov *et al.*, 1992; Imanshahidi and Hosseinzadeh, 2008), berberines, and protoberberines (e.g. tetrahydroberberine) (Karimov *et al.*, 1992) among which berberine, berbamine, jatrorrhizine, and palmatine are abundant in content. Alkaloids are undetectable in fruits and leaves (Koncic *et al.*, 2010a). Biphenyls and triterpens were largely isolated from twigs (Kim *et al.*, 2009; Kim, Choi, and Lee, 2010). The major chemical constituents of fruits are polyphenols including flavonoids (anthocyanins, flavonols, and flavanols), condensed and hydrolysable tannins, stibenoids (resveratrol), phenolic acids, and phenylpropanoid (Vereskovskii and Shapiro, 1986; Seeram, 2008). Flavonoids also could be isolated from the leaves (Imanshahidi and Hosseinzadeh, 2008).

The yield of crude methanol extract and individual compounds varied among plant organs. Koncic *et al.* (2010b) reported that the methanol extract was highest in fruits of *B. vulgaris* and *B. croatica* followed by leaves and lowest in roots or twigs, and berberine was largely found in twigs and roots while negligibly in leaves and fruits. Lv, Wang, and Xiao (1999) found that the amount of seven alkaloids (oxyacanthine,

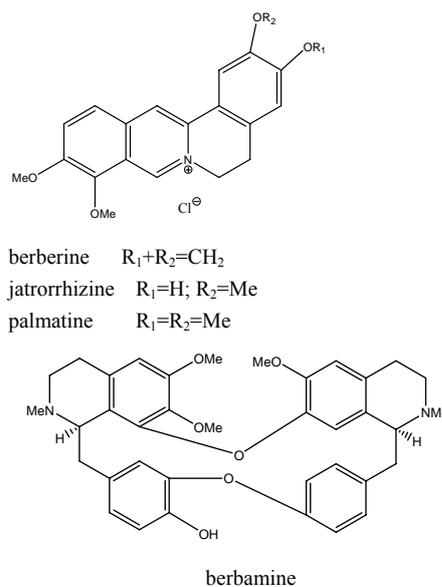


Fig. 1 Four major alkaloids in plants of *Berberis* L.

berbamine, isotetrandrine, columbamine, jatrorrhizine, palmatine, and berberine) from 17 species of *Berberis* L. was the highest in root barks followed by stem barks, root wood, and stem wood. In most roots of the 17 species, berberine was the richest alkaloid while berbamine, jatrorrhizine, and palmatine were irregular in content. Habitat-dependent variations in berberine content have been investigated (Andola *et al.*, 2010). The berberine content was significantly higher in *B. asiatica* Roxb. ex. DC. at low altitude than at high altitude, which was significantly influenced by the moisture and potassium percentage of the soil (Andola *et al.*, 2010).

Bioactive-guided isolation of bioactive compounds resulted in discovery of many nonalkaloids and alkaloids in this genus (Figs. 2–5). Recent studies showed that biphenyls and triterpenoids were isolated and identified from twigs of *B. kareana*, neither of which has been isolated from Berberidaceae (Kim *et al.*, 2009; Kim, Choi, and Lee, 2010). Berbekorin, a novel biphenyl, exhibited cytotoxic activity against the SK-MEL-2 skin melanoma cell line, other biphenyls (e.g. methoxyaucuparin, 3,5-dimethoxybiphenyl-2'-ol, 2'-hydroxy-3,4,5-trimethoxybiphenyl, and ϵ -cotonefuran) had inhibitory activities against NO production in LPS-activated microglial cell line (BV-2) (Kim *et al.*, 2009). Two triterpenoids, betulinic acid 3- β -*trans*-caffeate and betulinic acid 3- β -*cis*-caffeate, showed cytotoxicity against A594, SK-OV-3, SK-MEL-2, and HCT-15 cell lines (Kim, Choi, and Lee, 2010). 1-Methyl

Table 1 Chemical constituents in plants of *Berberis* L. discovered from 2008 to 2011

Compounds	Representative species	Parts	References
Alkaloids			
Proaporphine-type			
pronuciferine <i>N</i> -oxide	<i>B. coletioides</i>	whole	Fajardo <i>et al.</i> , 2009
pronuciferine	<i>B. coletioides</i>	whole	Fajardo <i>et al.</i> , 2009
Bisbenzylisoquinoline-type			
tabienine	<i>B. tabiensis</i>	stems	Quevedo <i>et al.</i> , 2008
Benzytetrahydroisoquinoline-type			
thalicmidine	<i>B. vulgris</i>	leaves	Imanshahidi and Hosseinzadeh, 2008
Berberine-type			
berlambine	<i>B. vulgris</i>	roots	Imanshahidi and Hosseinzadeh, 2008
Biphenyls			
berbekorin	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
2'-hydroxy-3,4,5-trimethoxybiphenyl	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
4,5-dihydroxy-3-methoxybiphenyl	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
eriobofuran	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
methoxyaucuparin	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
3,5-dimethoxybiphenyl-2'-ol	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
3-hydroxy-5-methoxybiphenyl	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
aucuparin	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
3,5-dimethoxybiphenyl	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
3,5-dimethoxybiphenyl-4'-ol	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
2'-hydroxyaucuparin	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
cotonefuran	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
9-hydroxyeriobofuran	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
cotonefuran	<i>B. koreana</i>	trunks	Kim <i>et al.</i> , 2009
Triterpenoids			
23- <i>trans-p</i> -coumaroyloxy-2a,3a-digydroxy-olean-12-en-28-oic acid	<i>B. koreana</i>	trunks	Kim, Choi, and Lee, 2010
23- <i>cis-p</i> -coumaroyloxy-2a,3a-digydroxy-olean-12-en-28-oic acid	<i>B. koreana</i>	trunks	Kim, Choi, and Lee, 2010
3b- <i>trans</i> -caffeoyloxy-2a-hydroxyurs-12-en-28-oic acid	<i>B. koreana</i>	trunks	Kim, Choi, and Lee, 2010
3- <i>O-trans-p</i> -coumaroyltormentic acid	<i>B. koreana</i>	trunks	Kim, Choi, and Lee, 2010
betulinic acid 3b- <i>trans</i> -caffeate	<i>B. koreana</i>	trunks	Kim, Choi, and Lee, 2010
betulinic acid 3b- <i>cis</i> -caffeate	<i>B. koreana</i>	trunks	Kim, Choi, and Lee, 2010
betulinic acid	<i>B. koreana</i>	trunks	Kim, Choi, and Lee, 2010
3-epimaslinic acid	<i>B. koreana</i>	trunks	Kim, Choi, and Lee, 2010
pomolic acid	<i>B. koreana</i>	trunks	Kim, Choi, and Lee, 2010
ursolic acid	<i>B. vulgris</i>	fruits	Imanshahidi and Hosseinzadeh, 2008
Flavonoid			
chrysanthemim	<i>B. vulgris</i>	fruits	Imanshahidi and Hosseinzadeh, 2008
delphinidin-3- <i>O</i> - β - <i>D</i> -glucoside	<i>B. vulgris</i>	leaves	Imanshahidi and Hosseinzadeh, 2008
pelargonin	<i>B. vulgris</i>	fruits	Imanshahidi and Hosseinzadeh, 2008
petunidin-3- <i>O</i> - β - <i>D</i> -glucoside	<i>B. vulgris</i>	fruits	Imanshahidi and Hosseinzadeh, 2008
Flavonol			
hyperoside	<i>B. vulgris</i>	fruits leaves	Imanshahidi and Hosseinzadeh, 2008

(To be continued)

(Continued Table 1)

Compounds	Representative species	Parts	References
quercitrin	<i>B. vulgris</i>	leaves	Imanshahidi and Hosseinzadeh, 2008
Polyphenols			
delphinidin-3-glucoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
cyanidin-3-glucoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
petunidin-glucoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
peonidin-3-glucoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
malvidin-3-glucoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
delphinidin-3-rutinoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
cyanidin-3-rutinoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
petunidin-3-rutinoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
peonidin-3-rutinoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
malvidin-3-rutinoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
delphinidin-3,5-dihexoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
cyanidin-3,5-dihexoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
petunidin-3,5-dihexoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
peonidin-3,5-dihexoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
malvidin-3,5-dihexoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
delphinidin-3-rutinoside-5-glucoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
petunidin-3-rutinoside-5-glucoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
malvidin-3-rutinoside-5-glucoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
delphinidin-3-sambubioside-5-glucoside	<i>B. microphylla</i>	fruits	Ruiz <i>et al.</i> , 2010
Phenylpropanoid			
caffeic acid	<i>B. vulgris</i>	fruit	Imanshahidi and Hosseinzadeh, 2008
chlorogenic acid	<i>B. vulgris</i>	fruit	Imanshahidi and Hosseinzadeh, 2008
Coumarin			
aesculetin	<i>B. vulgris</i>	fruit	Imanshahidi and Hosseinzadeh, 2008
Carbohydrate			
alpha-glucan	<i>B. vulgris</i>	leaf	Imanshahidi and Hosseinzadeh, 2008
pectin	<i>B. vulgris</i>	fruit	Imanshahidi and Hosseinzadeh, 2008
polysaccharide	<i>B. vulgris</i>	leaf	Imanshahidi and Hosseinzadeh, 2008
β-xylan	<i>B. vulgris</i>	leaf	Imanshahidi and Hosseinzadeh, 2008
sucrose	<i>B. vulgris</i>	fruit	Imanshahidi and Hosseinzadeh, 2008
vitamin			
ascorbic acid	<i>B. vulgris</i>	fruit leaf	Imanshahidi and Hosseinzadeh, 2008
vitamin K	<i>B. vulgris</i>	leaf	Imanshahidi and Hosseinzadeh, 2008

malate, a monoterpene, was isolated from the fruits of *B. integerrima* with enhancement of the antibacterial activity of Ampicillin (Alimirzaee *et al.*, 2009). 5'-Methoxyhydnocarpin-D and pheophorbide A isolated from the leaves of *B. fremontii* Torrey were identified as flavonolignan and porphyrin, respectively, which were inhibitors of multidrug resistant pump (MDR) in *Staphylococcus aureus* (Stermitz *et al.*, 2000b) (Fig. 5). In this study, the correct structure of

the first MDR inhibitor was determined as 5'-methoxy-hydnocarpin-D, a flavonolignan, instead of 5'-methoxy-hydnocarpin, an analogue of hydnocarpin reported previously (Stermitz *et al.*, 2000a). *O*-Methylisothalicberine, a bisbenzyl-isoquinoline alkaloid isolated from *B. chilensis* Gillies ex Hook, could significantly reduce mean arterial pressure in normotensive anaesthetized rats (Martinez, Torres, and Morales, 1997).

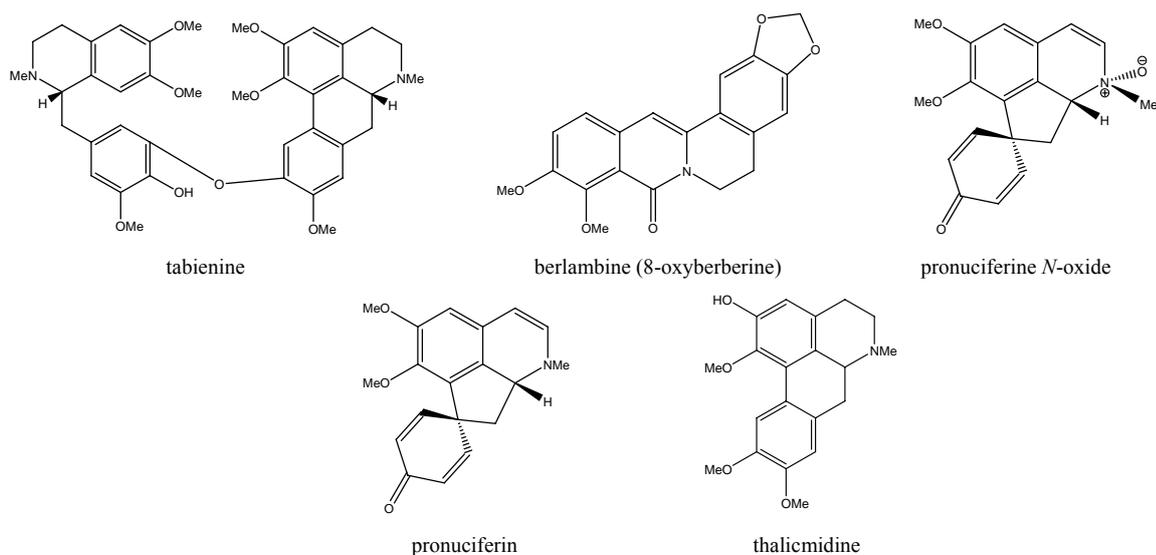


Fig. 2 Alkaloids in the genus *Berberis* L. discovered since 2008

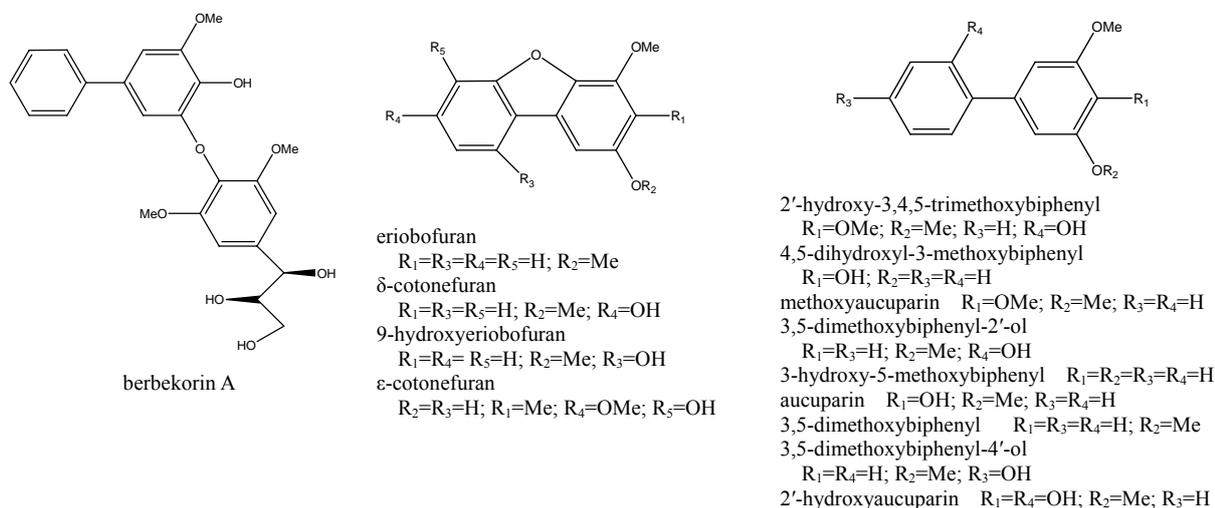


Fig. 3 Biphenyl compounds in plants of *Berberis* L.

Pharmacological activities of berberine and berbamine

Berberine and berbamine, the major components responsible for various bioactivities, have been intensively studied and reviewed (Tang *et al.*, 2009; Imanshahidi and Hosseinzadeh, 2008; Kulkarni and Dhir, 2010; Vuddanda, Chakraborty, and Singh, 2010). Here we summarized important potential bioactivities and the corresponding novel pharmacological mechanisms. Since berberine and berbamine are close in chemical structure—monoisoquinoline and bisbenzylisoquinoline respectively, they possess similarity and differences in several pharmacological activities, therapeutic effects, and molecular mechanisms.

Anticancer activity

Berberine and berbamine could induce apoptosis

in several tumor cell lines by loss in mitochondrial transmembrane potential and caspase activation, especially caspase-3 (Tang *et al.*, 2009; Wang *et al.*, 2007; 2009a; 2009b). They, however, possess difference and similarity in the mechanisms of apoptosis, cell-cycle arrest, cell-motility and invasion (Table 2).

Antitumor drugs are characterized by (1) promoting apoptosis including two pathways by activating caspase(s) cascade—Fas/FasL and mitochondria systems, Bcl-2, ICE, Apaf-1, c-myc, P53, ATM, and (2) inhibiting survival kinds of apoptosis e.g. PI3K/Akt, RAS-MAPK, ERK, NF- κ B, and NO signaling pathways, and the inhibitor of apoptosis (IAP) family of proteins (XIAP, CIAP1, CIAP2, Survivin) and/or fusion genes encoding oncoproteins. Berberine seems more likely to promote positive to apoptosis,

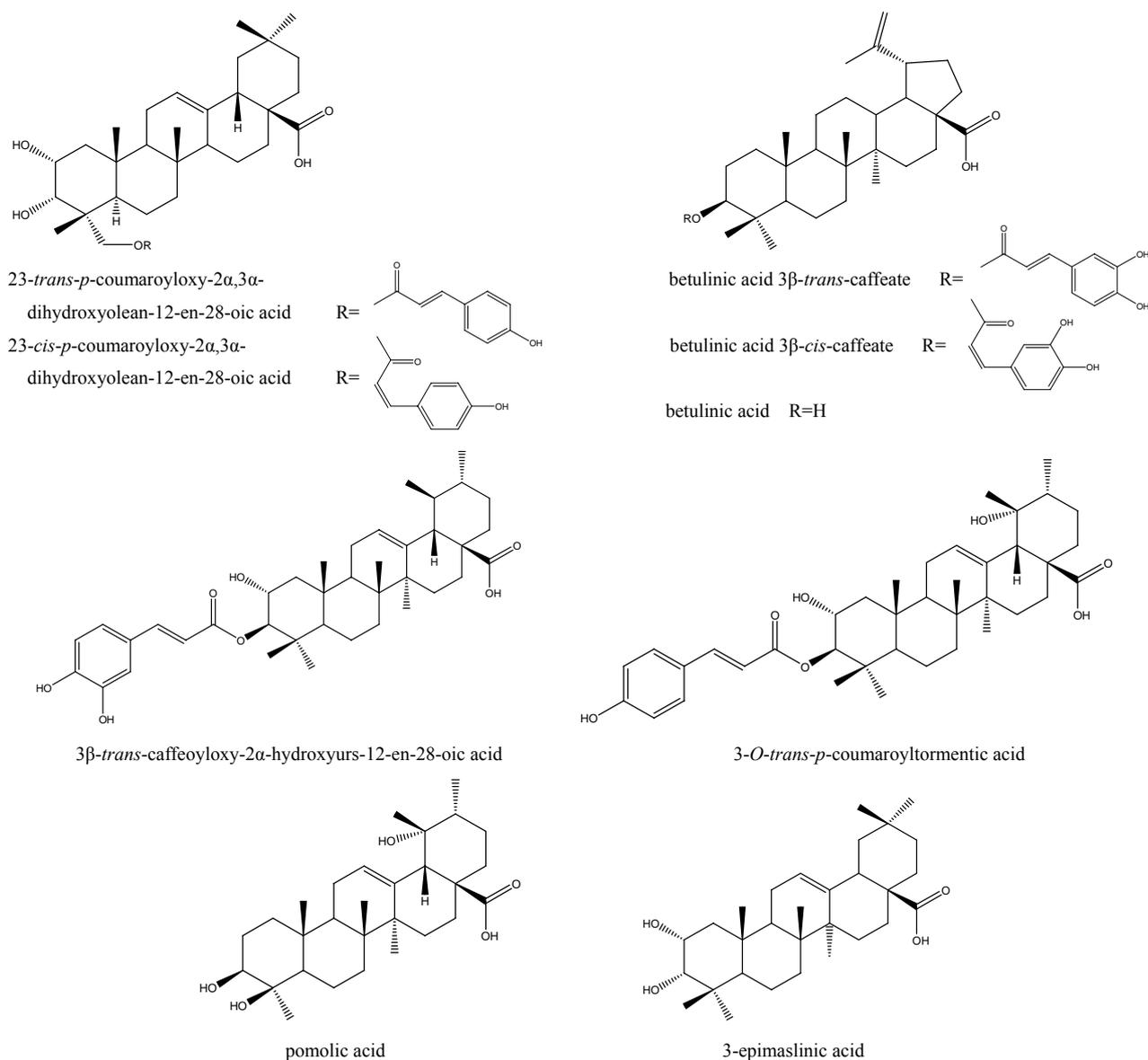
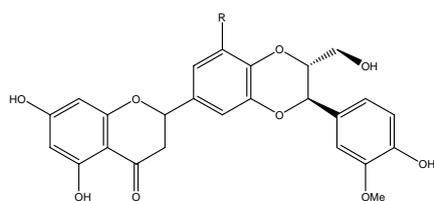


Fig. 4 Triterpenes in plants of *Berberis L.*

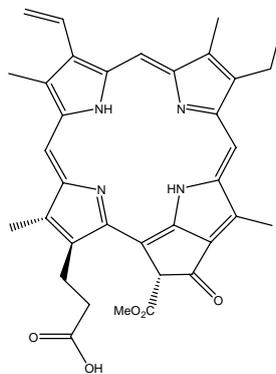
while berbamine and its analogues (e.g. 4-chlorobenzoyl berbamine) inclines to inhibit negatively to apoptosis or targeting oncogenes. They both induce Fas/FasL signaling pathways, activate P53, and alter ration of anti-apoptotic and pro-apoptotic members of the Bcl-2 family proteins (Wang *et al*, 2009a) in tumor cells. Berbamine and its analogues could induce apoptosis in tumor cells and multidrug resistant tumor cells by inhibiting PI3K/Akt and NF- κ B signaling pathways (Wang *et al*, 2009b; Liang *et al*, 2009; Du *et al*, 2010) and surviving gene's expression (He *et al*, 2006), such as *bcr/abl* fusion gene which induces leukemia-like syndrome *in vivo* (Xu *et al*, 2006; Wei *et al*, 2009b; Ji *et al*, 2002), *mdr-1* and P-GP (Dong *et al*, 2004; Han *et al*, 2003; Wei *et al*, 2009a; 2009b)—the genes and the

encoded protein that remove antitumor drugs out of tumor cells. In addition, Sun *et al* (2006) reported that berbamine reduced expression of Hsp90 protein, a molecular chaperone of p210 *bcr/abl* in chronic myeloid leukemia cells (Ph + K562) (Sun *et al*, 2006). Berbamine is also an inhibitor of telomerase which might play an important role in carcinogenesis and immortalization in human leukemia HL-60 cells (Ji *et al*, 2002).

Berberine and berbamine are able to induce the arrest in various phases of cell cycle. Berberine contributed to G₀/G₁, G₁, G₂, and G₂/M cell cycle arrests, and coordinated suppression of cyclin-dependent kinases (e.g., CDK 2, 4, and 6) and cyclins (e.g., cyclin B, D, and E) (Tang *et al*, 2009), while



5'-methoxyhydrocarpin-D



porphyrin

Fig. 5 Inhibitors of MDR isolated from *B. fremontii*

berbamine and its analogue (e.g. 4-chlorobenzoyl berbamine) induced cell-arrest at G_0/G_1 (Wang *et al.*, 2007), G_1 (Liang *et al.*, 2009), S (Dai *et al.*, 2009), and G_2/M (Liu *et al.*, 2010; Du *et al.*, 2010), and down-regulated cyclin D_1 (Liang *et al.*, 2009), B_1 and a cyclin-dependent kinase Cdc2/p34 (Liu *et al.*, 2010), cyclin A, B and a cyclin-dependent kinase CDK1 (Du *et al.*, 2010), which indicated *Sankezhen* could inhibit cell cycle at all phases.

Berberine and berbamine at low concentration could significantly inhibit migration ability of several human metastatic cancer cell lines by down-regulation of MMP proteins (e.g. MMP-1, MMP-2, and MMP-9) and some signaling pathways. Several studies showed that MAPK, NF- κ B, and RhoA signaling pathways were involved in berberine-treated cells (Ho *et al.*, 2009a; 2009b; Tsang *et al.*, 2009), while Ca^{2+} -dependent signaling pathway did in berbamine-treated ones (Huang *et al.*, 2010). In addition, berbamine could not only decrease MMP proteins but also increase TIMP-1 (Pan *et al.*, 2005), and berberine could decrease nuclear transcription factors e.g., c-fos, c-jun, AP-1 (a complex of c-fos and c-jun), and NF- κ B (Peng *et al.*, 2006; Fukuda *et al.*, 1999).

Both berberine and berbamine could inhibit cancer arising from leucocytes, liver, lung, stomach, colon, skin, oral, esophagus, brain, bone, and breast organs (Tang *et al.*, 2009; Duan *et al.*, 2009; Dai *et al.*, 2009;

Liang *et al.*, 2009; Wang *et al.*, 2009a) without toxicity against normal cells (Wang *et al.*, 2009b). However, they are different and similar likely in antitumor mechanism largely due to chemical structures that the former is monoisoquinoline and the latter is dimisoquinoline. It is primarily proposed that the two classes have tendency in opposite aspects of apoptosis, monoisoquinolines have tendency to positive aspect of apoptosis, while dimisoquinolines prefer to negative aspect of apoptosis which could integrate apoptosis. A recent study (Xie *et al.*, 2009) seems to support this proposal that berbamine derivatives are a class of compounds for antileukemia activity by inhibition of the cytoplasm-to-nucleus translocation of NF- κ B p65 which plays a critical role in the survival of leukemia stem cells. However, more evidences should be provided.

Immunopromotion and immunosuppression properties

Extracts of *Berberis* spp., such as *B. chengi* Chen, *B. lyceum* Royle, *B. asiatica* Roxb. ex DC., *B. crataegina* DC., and *B. koreana* (Cyran, 1989; Ikram, Ehsanul, and Warsi, 1966; Bhattarai, 1992; Yeşlada and Küpeli, 2002; Kim *et al.*, 2010), berberine and berbamine showed immunomodulative properties including up- and down-regulation.

Berberine chloride could ameliorate the spatial memory impairment and increase the expression of IL-1 β in improvement of Alzheimer's disease (Zhu and Qian, 2006). Berbamine is an immunopromotor in influenced mice, in which bioactivity of pulmonary macrophage is enhanced (Jin and Sui, 1986).

Berberine suppresses neuroinflammatory response through AMP kinase (AMPK) activation in BV-2 microglia, in which LPS- or interferon (IFN)- γ -induced iNOS and COX-2 expression is down-regulated (Lu *et al.*, 2010). In an LPS-injured rat model, berberine significantly suppressed the increased TNF- α , IL-1 β , and NO in plasma (Zhang *et al.*, 2011). Jeong *et al.* (2009) also reported that berberine suppressed proinflammatory response through AMPK activation in macrophages. Previous studies showed that berbamine had anti-inflammatory and immunosuppressive properties (Wong *et al.*, 1992; Küpeli *et al.*, 2002; Ren *et al.*, 2008). Ren *et al.* (2008) discovered that in EAE mice a novel immunomodulatory mechanism was induced by

Table 2 IC₅₀ values of berbamine and its derivatives against various tumor cell lines at 24, 48, and 72 h·μg⁻¹·mL⁻¹

Cells	Cell lines	IC ₅₀ / (?)	References
nasopharyngeal	KB	17.8	Marshall <i>et al.</i> , 1994
cervical	ES-2	13.67	Huang <i>et al.</i> , 2010
human hepatoma cell	SMMC7721	10.9**	Wang <i>et al.</i> , 2007
	DEE		Liu <i>et al.</i> , 2002
	HepG2	34.5*	Wang <i>et al.</i> , 2009a
	7402	1.167	Liu <i>et al.</i> , 2002
mouse hepatoma cell	H-22	3.312	Liu <i>et al.</i> , 2002
human oral squamous cell carcinoma			
chronic myelogenous leukemia	K562 (resistant)	11.1**	Wei <i>et al.</i> , 2009a
	K562	8**	Sun <i>et al.</i> , 2006
human leukemia cell	Gleevec-sensitive-K562	8.8	Xu <i>et al.</i> , 2006
	Gleevec-resistant-K562	11.34	Xu <i>et al.</i> , 2006
	KG-1	54.4	Xu <i>et al.</i> , 2006
	HL-60	15.0	Dong, Yang, and Kwan, 1997
	NL-60	< 10*	Ji <i>et al.</i> , 2002
	K562/Adr (resistant)	21.9	Dong <i>et al.</i> , 2004
human acute myelogenous leukemia	NB4	3.86**	Zhao <i>et al.</i> , 2007
	NB4	3.86	He <i>et al.</i> , 2006
	KM3	8.17	Liang <i>et al.</i> , 2009
human breast cancer cells	MCF-7	13.0	Dai <i>et al.</i> , 2009
	MDA-MB-231	13.7	Wang <i>et al.</i> , 2009b
	MDA-MB-435S	25	Wang <i>et al.</i> , 2009b
lymphoma cell line	Raji	0.9***	Du <i>et al.</i> , 2010
	L428	1.16***	Du <i>et al.</i> , 2010
	Namalwa	1.21***	Du <i>et al.</i> , 2010
	Jurkat	1.24***	Du <i>et al.</i> , 2010
human lung cancer	A549	< 20	Duan <i>et al.</i> , 2009
human myeloma cell	KM3	3.84	Liang <i>et al.</i> , 2009
human fibrosarcoma cells	HT1080	8.2	Pan <i>et al.</i> , 2005

*: IC₅₀ at 24 h; **: IC₅₀ at 48 h; ***: IC₅₀ of 4-chlorobenzoyl berbamine; others: IC₅₀ at 72 h

berbamine and probed its specific interactions with molecular targets, *vis.* Berbamine selectively inhibited IFN-γ through the Jak/Stat pathway in CD4⁺ T cells, in which berbamine selectively promoted STAT4 proteosomal degradation by up-regulating SLIM, a ubiquitin E3 ligase for STAT4.

Berberine and berbamine suppressed the delayed-type hypersensitivity (DTH) reaction response with sheep red blood cells (SRBC) (Ivanovska and Philipov, 1996; Luo *et al.*, 1998). Luo *et al.* (1998) demonstrated immunosuppressive effect of berbamine by DTH, the mixed lymphocyte reaction, and a skin

model of allograft rejection on mice, which indicated that berbamine could be a potential agent in clinical transplantation (Luo *et al.*, 1998).

Antidiabetic activity

Berberine has significant effect on carbohydrate metabolites, while no reports for berbamine. Recent preclinical and clinical studies (Zhang *et al.*, 2010; Kong *et al.*, 2009) showed it acted on glucose metabolism in rats and human of diabetes mellitus type 2 through several mechanisms, such as mimicking insulin, improving insulin action by activating AMPK, reducing insulin resistance through protein kinase

C-dependent up-regulation of insulin receptor expression, inducing glycolysis, and on increstin by promoting GLP-1 secretion, modulating its release, and inhibiting DPP-4 (Steriti, 2010). In addition, berberine has a protective effect for diabetes through β -cell regeneration, anti-oxidant enzyme activity, and decreasing lipid peroxidation (Vuddanda, Chakraborty, and Singh, 2010). Clinical studies showed that berberine was as effective as metformin (3×500 mg of each) in the bioactivities of reducing elevated blood glucose. Adverse effects of transient gastrointestinal were fairly common in berberine-administrated patients (Zhang *et al.*, 2010).

A recent study (Cui *et al.*, 2009) reported that berberine could ameliorate diabetes mellitus type 1 NOD mice by suppressing T cell differentiation. In this study, berberine inhibited Th17 differentiation by activating ERK 1/2 and inhibited Th1 differentiation by inhibiting p38 MAPK and JNK activation, and it controlled the stability of STAT4 through the ubiquitin-proteasome pathway. This finding revealed that the immune regulation property of berberine was another mechanism for type 1 diabetes treatment.

Antihyperlipidemic activity

Lipid metabolic effect of berberine has been observed in human and animals, while no reports for berbamine. The mechanisms included upregulation of low-density lipoprotein receptor (LDLR), promotion of LPL activities and adipose hormones (visfatin and adiponectin), and inhibition of lipid biosynthesis and pro-lipid cell differentiation (Shen *et al.*, 2010).

Berberine, only depending on ERK activation but sterol regulatory element binding protein (SREBP), elevates hepatic LDLP expression through a post-transcriptional mechanism that stabilizes the *LDLP* mRNA, and the responsible agent is the 5' proximal section (AU-rich elements and tetranucleotide UCAU) of the *LDLR* mRNA 3' untranslated region (3' UTR), which is a unique mechanism distinct from statins (Kong *et al.*, 2004). And *LCLR* mRNA stability is controlled by a group of AU-rich elements binding proteins including hnRNP D, hnRNP I, and KSRP. Berberine interferes with the binding proteins-receptor interaction, which is a likely mechanism for regulating LDLR expression (Li *et al.*, 2009). Berberine may be useful supplement to statins due to the following

reasons. It is time and dose-dependent to up-regulate hepatic *LDLR* mRNA as well as down-regulate proteins convertase subtilisin/kexin type 9 (PCSK9) which is benefit to LDLR degradation. However, statins up-regulate both *LDLR* mRNA and PCSK9 depending on SREBP, which would contract therapeutic effects of statins by reducing PCSK9 expression. A combination of berberine and mevastatin increases LDLR mRNA and protein levels, while suppressing the increase in PCSK9 mRNA levels caused by mevastatin alone (Cameron *et al.*, 2008). SP600125, the inhibitor of JNK, could suppress the berberine-induced up-regulation of LDLR mRNA because both ERK and JNK/c-jun are activated by berberine.

Berberine inhibits total cholesterol (TC) and triglyceride (TG) biosynthesis in HepG2 by activating of AMPK, and promotes oxidation of lipid acids as well. The underlying inhibitory mechanism is the same as AICAR—the activator of AMPK, and could be inhibited by PD98059—the inhibitor of MAPK/ERK (Shen *et al.*, 2010).

Berberine inhibites 3T3-L1 adipocyte differentiation through PPAR pathway, in which the expressions of PPAR α , β/δ , γ , and C/EBP α mRNA and proteins are inhibited. However, berberine up-regulates the expression of PPAR α mRNA in hepG2 (Shen *et al.*, 2010).

Antihypertensive activity

Previous studies demonstrated blood pressure lowering and vasorelaxatory effects of berberine and berbamine. For berberine, the mechanisms include α 1-adrenoceptor antagonistic action in rat and rabbit aorta, ACE-inhibitory activity and direct release of NO/cGMP from rat aortic rings, the activation of tetrapentylammonium, 4-aminopyridine, and Ba²⁺-sensitive K⁺ channels, the inhibition of intracellular Ca²⁺ release from caffeine-sensitive pools, the blocking of L-type calcium channels, the potentiation of acetylcholine (Imanshahidi and Hosseinzadeh, 2008), the inhibition of rat vascular smooth muscle cell proliferation and migration *in vitro*, and the promotion of neointima formation after balloon injury *in vivo* or in a rat model (Lee *et al.*, 2006). For berbamine, the major mechanism is blocking calcium channels (Guo and Fu, 2005). Han *et al.* (2006) reported that *N*-methyl berbamine increased calcium-activated potassium currents in the rat mesenteric resistance vascular smooth muscle cells, which could lead to lower blood pressure.

Anti-arrhythmic activity

Berberine and berbamine are both found with anti-arrhythmic activities. Berbamine is a popular drug of anti-arrhythm through inhibiting ionic channels of sodium, potassium, calcium, etc., negative frequency and negative transduction, improving the diastolic excitation threshold of myocardium, and prolonging effective refractory period of myocardium (Guo and Fu, 2005).

Recent studies showed that berberine which has been used in diabetic patients with ischemic cardiomyopathy might effectively reduce the morbidity and lethality of myocardial infarction related arrhythmias. Wang *et al* (2011) revealed that berberine could elicit this effect via $I_{K1}/Kir2.1$ in rat type 2 diabetic myocardial infarction model.

Anti-osteoporotic activity

Berberine by ig administration inhibited bone resorption of ovariectomized rats. And berberine inhibited osteoclast formation in coculture system of

mouse primary osteoclastic cells and bone narrow cells in the presence of $1\alpha, 25$ -dihydroxyvitamin D₃, parathyroid hormone, and interleukin- 1α (IL- 1α) (Li *et al*, 1999). Hu *et al* (2008) reported that berberine inhibited RANKL-induced osteoclast ad survival through suppressing the NF- κ B and Akt pathways. Berberine attenuated RANKL-induced activation of NF- κ B through inhibiting phosphorylation at the activation loop of I κ B α kinase β , phosphorylation and degradation of I κ B α , and NF- κ B p65 nuclear translocation.

Neuroprotective activity

Berberine possesses neuroprotective activities in various animal models of neurodegenerative and neuropsychiatric disorders such as Alzheimer's disease, forebrain ischemia, mental depression, anxiety, pentylenetetrazole, maximal electroshock, kainic acid-induced convulsion (Bhutada *et al*, 2010), and disorders associated with excitotoxicity (Campisi *et al*, 2010) (Table 3).

Table 3 Therapeutic use of berberine in various CNS disorders and the underlying mechanisms

Clinical disease and syndrome	Mechanisms	References
Alzheimer's	an increased level of both IL-1 and iNOS resulted in neuronal destabilization acetylcholinesterase inhibiting property cholesterol lowering agent	Kulkarni and Dhir, 2010
Schizophrenia	prolyl oligopeptidase inhibition D2 dopamine receptor antagonist property	
Mental depression	monoamine oxidase property norepinephrine, serotonin and dopamine transporter inhibitor modulatory effect on sigma receptors	
Cerebral ischemia	N-methyl-d-aspartate receptors antagonistic activity anti-inflammatory activity reduced hydrogen peroxide-induced free calcium concentration elevation	
Anxiety	modulate serotonergic system in the body	
Glutamate-evoked excitotoxicity	ameliorate excessive production of glutamate, protein misfolding and aggregation, mitochondrial fragmentation and neurodegeneration	Campisi <i>et al</i> , 2010
Pentylenetetrazole, maximal electroshock, and kainic acid-induced convulsion	modulate neurotransmitter systems	Bhutada <i>et al</i> , 2010

Anti-oxidative activity

Berberine and berbamine have been found to be anti-oxidants (Guo and Fu, 2005, Shirwaikar *et al*, 2006). Thirupurasundari, Padmini, and Devaraj (2009)

reported that berberine significantly attenuated the increase on lipid peroxidation and protein bound carbohydrates and enhanced the anti-oxidative status in azoxymethane-induced colon cancer in rats, which

indicated that berberine inhibited neoplastic transformation by the induction of anti-oxidant defence system and ability to induce apoptosis. Berbamine was also an anti-oxidant which scavenged O_2^- in alkaline DMSO and xanthine/xanthine oxidase systems, respectively, in mice livers (Ju and Han, 1990). Zhang, Ba, and Ren (1993) reported that berbamine could increase SOD and GSH-Px activities and reduce lipid peroxidation and O_2^- in rabbits with myocardial infarction. Zhou *et al.* (1998) demonstrated that berbamine reduced O_2^- in rats with cerebral ischemia. Extracts of *Berberis* spp. (e.g. *B. aristata*, *B. vulgaris*, *B. croatica*, and *B. microphylla*) had anti-oxidative activities (Singh and Kakkar, 2009; Koncic *et al.*, 2010b; Hanachi, 2009; Ruiz *et al.*, 2010), some of which were demonstrated to correlate well with the content of phenols and flavonols (Koncic *et al.*, 2010b; Ruiz *et al.*, 2010).

Discussion

Determination of botanical origins

Botanical origin is the first important issue in development of quality assessment of *Sankezhen*. As we know, *Sankezhen* has wide botanical origins which spread worldwide. It is essential to determine the representative species, the corresponding producing areas, and the harvest periods which certainly will contribute to variations in chemical composition and contents. As a result, the quality of *Sankezhen* is approaching controllable, the ideal botanical origins are characterized by high yield (e.g. population biomass and desired compound contents) and stable genetics, which should be further investigated and determined.

Quality and quantity of alkaloids in various parts of *Berberis* spp.

Chemical constituents in the representative species should be intensively isolated and identified, which is the base for quality control and pharmaceutical effects.

Xiao *et al.* (1974) qualitatively and quantitatively determined berberine, palmatine, jatrorrhizine, and berbamine from over 40 species by TLC and spectrophotometry. Later studies were focused upon the analysis of contents of those alkaloids in *Berberis* spp. from different habitats and harvest periods, different species and the various parts by HPLC in China, aiming to find out alkaloid distribution in various parts of

plants and content variations in different conditions (Lv, Wang, and Xiao, 1999; Yang, Ma, and Qi, 2009). Few studies have been intensively carried out upon chemical compounds of the species of *Berberis* L. in China, and only Wang *et al.* (2009c) investigated the chemical constituents in *B. sargentiana*. Moreover, several studies (Kim *et al.*, 2009; Kim, Choi, and Lee, 2010) have demonstrated that plants in *Berberis* L. species produced various interesting chemical structures besides the four common alkaloids (see the last paragraph). Thus, we believe that it is very valuable to systematically study chemical components from each species, especially the representatives and the four species admitted in *Chinese Pharmacopeia 2010*. Obviously chemical constituents in detail are quite necessary in the establishment of quality control methods of great specialization. In addition, it is suggested to investigate the certain producing areas and the harvest periods since they greatly affect the contents of chemical compounds, especially for such a herb as *Sankezhen* with rather wide distribution in China.

Development of authentication approaches

Chemical constituents, bioactivities, and botanical origin are the important factors in authentication of a Chinese materia medica. Berberine is the only chemical marker for quality assessment of *Sankezhen* in *Chinese Pharmacopeia 2010*, which is the single-compound analysis approach (Yuan, Zhang, and Xiao, 2011). However, simultaneous analysis on four alkaloids including berberine, berbamine, jatrorrhizine, and palmatine by HPLC-DAD has been reported in several papers (Di *et al.*, 2004; Lv, Wang, and Xiao, 1999). HPLC simultaneous analysis on the four chemical markers is increasingly popular in recent years, which is the multi-compound analysis approach (Yuan, Zhang, and Xiao, 2011). HPLC spectrum of the four alkaloids will be more accurate in authentication than one. Sometimes it is difficult to distinguish *Sankezhen* and *Huangbai* due to similarity in spiny. As a result, folk people mis-submit *Sankezhen* as *Huangbai* so multi-compound analysis approach will be a method of more accurate and feasible in identification. Thus HPLC simultaneous determination of four markers might be used for assessing quality of *Sankezhen*.

Obviously other authentication approaches need to be developed to overcome the limitation of each

approach. Bioassay approach is a new quality control method for Chinese materia medica, which shows the advantages in case of uncertainty of chemical compounds and is expected to be well developed in the Guide of *Chinese Pharmacopoeia 2010*. This approach addresses the biological properties which closely relate to efficacy and safety. Yan *et al* (2008) assayed antibacterial potency of berberine alkaloids in *Coptis Rhizoma* using microcalorimetry and thermoanalysis, which is a promising method in quality assessment.

DNA-based markers and genomics are the promising methods for authentication of medicinal plants (Hao *et al*, 2009). And DNA barcoding—a method that is the use of short DNA sequences for species identification (Han *et al*, 2010), is expected to be accepted in updating *Chinese Pharmacopoeia* due to the advantages of quickness and accuracy. However, it is doubted in the genus of *Berberis* L. because universal plant DNA barcode loci may not work in a complex genus (Roy *et al*, 2010). This is a result obtained from Indian berberis samples, which is not fully convinced in Chinese berberis. Nevertheless, it is quite hard to identify origin plants of *Berberis* L. on market. Therefore, proper methods of DNA-based markers should be established.

Pharmaceutical and therapeutic effects of Sankezhen—a prospect natural resource for drug discovery

Sankezhen has wide potential therapeutic effects on diseases from cardiovascular, immune, central nervous, respiratory, and several stubborn diseases such as cancer, diabetes, Alzheimer's disease, etc. However, some of the pharmaceutical effects are uncertain in clinical practice, which indicates that there is a long way from discovery of bioactivities to clinical practice. Fortunately, there is an example for *Sankezhen* studies, that is *Huanglian* (*Coptidis Rhizoma*) and berberine, which have been quite intensively studied and recognized as novel antineoplastic agents (Tang *et al*, 2009), and *Sankezhen* is a substitute of *Huanglian* in TCM to treat gastrointestinal diseases. Nevertheless, we do not suppose that *Sankezhen* is identical to *Huanglian* in all pharmaceutical effects. Since few studies are available in identical property, the two herbs are suggested to be studied in identities and differences including chemical constituents, pharmaceutical

bioactivities, and ethnopharmacology (e.g. nature, flavour, and favored channel tropisms).

Other parts of *Berberis* spp. and active compounds

Twigs, trunks, and barks are bioactive parts of *Berberis* spp. due to accumulation of alkaloids (Table 1), however they are unable to substitute the roots unless clinical efficacy evidences are provided. New classes of chemical compounds (e.g. triterpenes and biphenyls from trunks of *B. koreana*) (Kim *et al*, 2009) and novel compounds from the extract of those parts of the species of *Berberis* L. have been isolated and identified by bioactivity-guided assays, which indicated that this genus was a great reservoir with potential wide spectrum therapeutical effects. Triterpenes from trunks of *B. koreana* are characterized by five rings including oleanane, ursane, and lupine, which are known in antitumor bioactivities against various cancer cell lines (Sun and Li, 2009). Biphenyls are another class of compounds isolated from *Berberis* L. with several substituent groups of -OCH₃ and OH, which indicates the bioactivities of depressing center central nervous system and relaxing muscles due to similar chemical structures with magnolol. We suppose that diversity of chemical groups contributes to multispectrum bioactivities of *Berberis* spp. properly in some ways different from alkaloids. However, further studies are needed upon bioassays of those compounds from the plants of *Berberis* L.

Fruits are also the important pharmaceutical part of *Berberis* spp. in some regions or ethnic minorities which are promising pharmaceutical parts and deserve further studies, e.g., in China, the fruits of *B. heteropoda* Schrenk, neutral in nature and invigorating the gastrointestinal tract and clearing away heat and toxic material, are authenticated as the botanical origin in Ha ethnopharmacology to treat gastroenteritis, dysentery, dyspepsia, aphthous stomatitis, and vitamin E deficiency disease (Ha and Xv, 2005). Fruits of *B. henryana* Schneid are used to treat infantile convulsion, release wind and fire from inner body in ethnic minority areas of Sichuan Province in China (Xiao *et al*, 1974). In Croatia, fruits of *B. croatica* and *B. vulgaris* are used for ailments and discomforts of the kidney and urinary tract, gastrointestinal tract, liver disease, bronchial discomfort, spleen ailments, spasms, and as a stimulant for the circulatory system (Koncic *et al*,

2010b). *B. vulgaris* fruit is known for its anti-arrhythmic and sedative effects in Iranian traditional medicine (Fatehi *et al.*, 2005). A few papers reported that chemical constituents of fruits were polyphenols including flavonoids (anthocyanins, flavonols, and flavanols), condensed and hydrolysable tannins, stibenoids (resveratrol), phenolic acids, and phenylpropanoid. However, fruits of *Berberis* spp. in China have never been systematically or intensively studied on chemical constituents and biological activities, which needs to be carried out for the discovery of new botanical resources as crude drugs.

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