

Genus *Gentianella* Moench: A Phytochemical and Ethnopharmacological Review

LI Min-hui^{1*}, LI Li^{2,3}, YANG Yu-mei¹, ZHANG Na¹, SONG Xiao-ling¹, XIAO Pei-gen^{2,3*}

1. Baotou Medical College, Baotou 014060, China

2. School of Chinese Materia Medica, Beijing University of Chinese Medicine, Beijing 100102, China

3. Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences, Beijing 100193, China

Abstract: The plants in genus *Gentianella* Moench (Gentianaceae) which comprised approximately 250 species, are mainly distributed in temperate regions of the world. Many *Gentianella* plants are intensely bitter and employed in traditional medicine to stimulate appetite, treat disorders of the gallbladder, and treat fever like the other bitter gentians in various regions of the world. Some species exhibit other remarkable therapeutic effects in the treatments of obesity, diabetes, and heart diseases. Eleven iridoids, twenty-eight xanthones, three C-glucoflavonoids, and eight other compounds have been isolated from the genus. Most of these compounds are associated with antimicrobial, anti-inflammatory, anti-oxidant, hypoglycemic, and antitumor activities, which provide an empirical base for the traditional utilization of the plants in genus *Gentianella* Moench.

Key words: ethnopharmacology; *Gentianella* Moench; pharmacology; phytochemistry; traditional medicine

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Introduction

As natural drugs now play an even more important role in medical and healthcare services, the ethnopharmacology of medicinal plants has attracted increasing attention in new drug research and development.

The plants of *Gentianella* Moench (Gentianaceae) mainly distributes in temperate regions of the world. It comprises approximately 250 annual, biennial or perennial species with 4 or 5-merous flowers, obconical or cylindrical corolla which is fimbriate in the throat and with erect or patent lobes, no style, but sessile and stipitate ovary (Von Hagen and Kadereit, 2001). The group is an extremely abundant medicinal plant source. Many species of the genus have traditional remedies for loss of appetite, jaundice, disorders of the gallbladder, fever, etc. In recent years, extracts of some *Gentianella* Moench species have been also patented for improving insulin resistance, liver function, and decreasing blood sugar, uric acid, and obesity in US and Japan (Ogawa *et al*, 2006; Ogawa, 2006; 2007).

Previous phytochemical investigations on the plants of *Gentianella* Moench have resulted in the isolation of xanthones, C-glucoflavones, and terpenoids (iridoids, sesterterpenoids, and triterpenes). The occurrence of iridoids, xanthones, and C-glucoflavones is typical of genus *Gentianella* Moench (Jensen and Schripsema, 2002; Janković, Krstić, and Ivana, 2005), and the medicinal value of the genus is just due to presence of these compounds. Some plants of *Gentianella* Moench are either very rich in these compounds or have been studied extensively. For example, 14 different compounds have been isolated from *G. nitida* (Grisebach) Fabris (Lacaille-Dubois and Galle, 1996; Kawahara *et al*, 1997; Kawahara, Nozawa, and Kurata, 1999; Kawahara, Nozawa, and Sekita, 2001.).

Recently more and more researchers have increasing interest on the species of *Gentianella* Moench and their biological activities. In this review, the phytochemical and pharmacological research on the species of *Gentianella* Moench together with their traditional uses will be presented

* Corresponding author: Li MH Address: Baotou Medical College, Baotou, Inner Mongolia 014060, China
Tel/Fax: +86-472-7167 795 E-mail: li_minhui@yahoo.cn

Xiao PG Address: Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences, Beijing 100193, China
Tel/Fax: +86-10-6281 8235 E-mail: xiaopg@public.bta.net.cn

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and critically evaluated.

Traditional medicinal uses

Many plants of *Gentianella* Moench are intensely bitter and employed in traditional medicines to stimulate appetite, treat disorders of the gallbladder and treat fever like the other bitter gentians in various regions of the world, such as *G. florida* (Griseb.) Holub, *G. achalensis* (Gilg.) T. N. Ho et S. W. Liu, *G. austriaca* (A. et J. Kerner) Holub, *G. amarelle* ssp. *acuta* (Michx.) J. M. Gillett, etc (Ratera and Ratera, 1980; Nadinic et al, 1997; 1999; Vinterhalter et al, 2008; Urbain et al, 2008). Some species exhibit other remarked therapeutic effects in the treatments of obesity, diabetes, and heart diseases (Ma, 1993; Kawahara et al, 1997; Kawahara, Nozawa, and Flores, 2000).

Some representative medicinal species of *Gentianella* Moench are listed:

G. nitida, a biennial medicinal plant growing in the Andes region, is used in traditional Peruvian folk medicine (Commonly known as “Hercampuri” or “Hircampure”) as a cholagogue and for the treatment of hepatitis and obesity (Kawahara et al, 1997; Kawahara, Nozawa, and Kurata, 1999; Kawahara, Nozawa, and Sekita, 2000; Kawahara, Nozawa, and Sekita, 2001).

G. achalensis is a small herb distributed in the northwestern and central regions of Argentina. It is locally known as “pasto amargo” (bitter grass). The aerial parts of the plant have been extensively used for digestive and liver problems (Nadinic et al, 1999).

G. alborosea (Gilg.) Fabris is a herb which grows in the Peruvian highlands. It has been used since the times of the Incas to relieve stomach pain, regenerate hepatic functions, fight fever produced by malaria, detoxify blood, regulate circulation, and reduce obesity (Kawahara, Nozawa, and Flores, 2000; Acero, Llinares, and Galán, 2006). In addition, the plant is also employed traditionally in acne treatment (Rainer, Bussmann, and Doris, 2008).

G. acuta (Michx.) Hulten, locally known as *Guixincao*, has been used as herb tea with the health properties of clearing heat and toxic materials, removing pathogenic heat from blood, and increasing secretion of urine in the eastern region of Inner Mongolia (China). In addition, the Owenke (one of 56 ethnic minorities in China) use the aerial parts of *G. acuta* for treating heart diseases (Ma, 1993).

G. nevadensis (Gilg.) Weaver et Rudenberg is known as “dictamo real” in Venezuela. A decoction or an alcoholic beverage of the roots and aerial parts of this plant was employed in folk to relieve pain, regulate the circulation, and promote digestion (Báez, Quintero, and Nieves, 1999).

G. azurea (Bunge) Holub is an annual herb distributed in West China. It is used in Tibetan traditional medicine for clearing heat and drying dampness. The frequently clinical application of the plant has been for the treatment of fever and disorders of the gallbladder (Zhang and Yang, 1994).

In addition, *G. bicolor* (Wedd.) J. S. Pringle, *G. crassicaulis* J. S. Pringle, *G. dianthoides* (Kunth) Fabris ex J. S. Pringle, *G. graminea* (Kunth) Fabris, and *G. thyrsoidae* (Hook.) Fabris are used to treat diabetes and hyperlipidemia in South America (Tomas, Lock, and Jurupe, 1999; José et al, 2005; Rainer and Bussmann, 2006). *G. bruneotricha* (Gilg.) J. S. Pringle, known as “anga macha” in Northern Peru, is employed in folk to treat postnatal infection (Rainer and Bussmann, 2006).

Phytochemistry

Iridoids

Iridoids, a group of monoterpenoid compounds with a cyclopentanopyran ring structure, are the most common and characteristic compounds in the family Gentianaceae. The majority of iridoids found in nature are in the form of glucosides, and the iridoid glucosides in the plants of *Gentianella* Moench are usually seco-iridoids. The distribution and the structures of iridoids in the plants of *Gentianella* Moench are shown in Table 1 and Fig. 1.

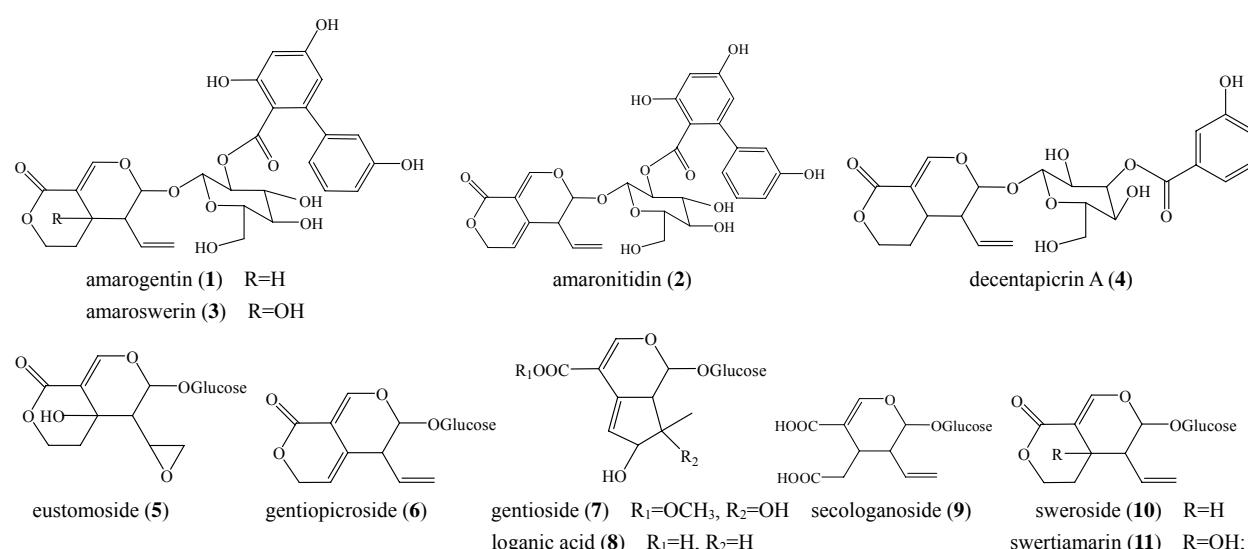
Xanthones

Xanthones are a class of secondary metabolites with a variety of structures in oxidation pattern. Most xanthones are mono- or poly-methyl ethers or are found as glycosides. They have a rather restricted occurrence among higher plants and have been used as chemotaxonomic markers at infra- and super-genus levels in the family Gentianaceae (Massias, Carbonnier, and Molho, 1982). The known occurrences of xanthones in the plants of *Gentianella* Moench are listed in Table 2. The structures of xanthones in the genus are shown in Fig. 2.

There is an evidence that 1,3,5-trihydroxyxanthone (a) and 1,3,7-trihydroxyxanthone (b) represent the primitive

Table 1 Iridoids in plants of *Gentianella Moench*

| Compounds | Species | References |
|---------------------|---|---|
| amarogenin (1) | <i>G. alborosea</i> <i>G. multicaulis</i> (Gillies ex Griseb.) Fabris | Singh, 2008 Rosella et al, 2007 |
| amaronitidin (2) | <i>G. nitida</i> | Lacaille-Dubois and Galle, 1996 |
| amaroswerin (3) | <i>G. nitida</i> | Kawahara N, Nozawa M, and Sekita, 2001 Lacaille-Dubois and Galle, 1996 |
| decentapicrin A (4) | <i>G. nitida</i> | Kawahara N, Nozawa M, and Sekita, 2001 |
| eustomoside (5) | <i>G. bulgarica</i> (Velen.) Holub | Do et al, 1987 |
| gentiopicroside (6) | <i>G. acuta</i> <i>G. amarella</i> ssp. <i>acuta</i> <i>G. azurea</i> <i>G. bulgarica</i> <i>G. campestris</i> (L.) Boerner | Lv and Li, 2009 Urbain et al, 2009 Zhang and Yang, 1994 Do et al, 1987 Urbain et al, 2009 |
| gentioside (7) | <i>G. ramosa</i> (Hegetschw.) Holub | Rosella et al, 2007 Urbain et al, 2009 |
| loganic acid (8) | <i>G. bulgarica</i> | Do et al, 1987 |
| secologanoside (9) | <i>G. nitida</i> | Zhang and Yang, 1994 Lacaille-Dubois and Galle, 1996 |
| sweroside (10) | <i>G. bulgarica</i> <i>G. multicaulis</i> | Do et al, 1987 Rosella et al, 2007 |
| swertiaarin (11) | <i>G. parviflora</i> (Griseb.) T. N. Ho. <i>G. acuta</i> <i>G. azurea</i> <i>G. bulgarica</i> <i>G. parviflora</i> | Rosella et al, 2007 Lv and Li, 2009 Zhang and Yang, 1994 Do et al, 1987 Rosella et al, 2007 |

**Fig. 1 Structures of iridoids in plants of *Gentianella Moench***

state in the two different biosynthetic pathways (Jensen and Schripsema, 2002). The 1,3,5,8- and 1,3,7,8-tetra-substituted xanthones are formed by an oxidation at position 8 of either the basic structures a or b (Jensen and Schripsema, 2002; Lv and Li, 2009). According to the data in the Table 2 and Fig. 2, the predominant oxidation pattern of the xanthones found in the plants of *Gentianella Moench* so far is 1,3,5,8-, and to a lesser extent, 1,3,7,8-substitution.

C-glucoflavonoids

C-glucoflavonoids, in which the sugar is attached

to the flavonoid nucleus by a carbon-carbon bond, have limited distribution in the family Gentianaceae in comparison with iridoids and other xanthones (Jensen and Schripsema, 2002; Janković, Krstić, and Ivan, 2005), but are found commonly available in the plants of *Gentianella Moench* (Table 3 and Fig. 3).

Others

About eight other compounds, including two pentacyclic triterpenes (oleanolic acid 43 and ursolic acid 44) (Zhang and Yang, 1994; Nadinic et al, 1997; 1999; Tomas, Lock, and Jurupe, 1999; Lv and Li, 2009), two

Table 2 Xanthones in plants of *Gentianella* Moench

| Compounds | Species | References |
|---|---|---|
| 1-hydroxy-3,5,8-trimethoxyxanthone (12) | <i>G. thyrsoidae</i> | Tomas, Lock, and Jurupe, 1999 |
| 1-hydroxy-3,6,7,8-tetramethoxy-xanthone (13) | <i>G. thyrsoidae</i> | Tomas, Lock, and Jurupe, 1999 |
| 1,5,8-trihydroxy-3,4-dimethoxy-xanthone (14) | <i>G. parviflora</i> | Rosella et al, 2007 |
| 1,7,8-trihydroxy-3,4-dimethoxy-xanthone (15) | <i>G. parviflora</i> | Rosella et al, 2007 |
| 3-methylcorymbiferin (16) | <i>G. antarctica</i> (Kirk) T.N.Ho et S.W.Liu <i>G. cerina</i> (Hook.f.) T.N.Ho et S.W.Liu <i>G. corymbifera</i> (Kirk) Holub <i>G. patula</i> (Kirk) Holub | Benn et al, 2009 Benn et al, 2009 Benn et al, 2009 Benn et al, 2009 |
| bellidifolin (17) | <i>G. achalensis</i> <i>G. acuta</i> <i>G. albanica</i> (Javorka) Holub <i>G. amarella</i> ssp. <i>acuta</i> <i>G. antarctica</i> <i>G. azurea</i> <i>G. bellidifolia</i> (Hook.f.) Holub <i>G. bulgarica</i> <i>G. campestris</i> <i>G. cerina</i> <i>G. corymbifera</i> <i>G. crispatia</i> <i>G. florida</i> <i>G. multicaulis</i> <i>G. nevadensis</i> <i>G. patula</i> <i>G. parviflora</i> <i>G. ramosa</i> <i>G. serotina</i> (Cockayne) T.N.Ho et S. W. Liu <i>G. thyrsoidae</i> | Nadinic, Penna, and Saavedra, 2002 Lv and Li, 2009 Janković, Krstić, and Ivana, 2005 Urbain et al, 2009 Benn et al, 2009 Zhang and Yang, 1994 Benn et al, 2009 Janković, Krstić, and Ivana, 2005 Urbain et al, 2009 Benn et al, 2009 Benn et al, 2009 Janković, Krstić, and Ivana, 2005 Nadinic et al, 1997 Rosella et al, 2007 Báez, Quintero, and Nieves, 1999 Benn et al, 2009 María et al, 2007 Urbain et al, 2009 Massias, Carbonnier, and Molho, 1982 Tomas, Lock, and Jurupe, 1999 |
| corymbiferin (18) | <i>G. antarctica</i> <i>G. albanica</i> <i>G. bellidifolia</i> <i>G. bulgarica</i> <i>G. cerina</i> <i>G. corymbifera</i> <i>G. crispatia</i> <i>G. patula</i> <i>G. albanica</i> | Benn et al, 2009 Janković, Krstić, and Ivana, 2005 Benn et al, 2009 Janković, Krstić, and Ivana, 2005 Benn et al, 2009 Benn et al, 2009 Janković, Krstić, and Ivana, 2005 Benn et al, 2009 Benn et al, 2009 Janković, Krstić, and Ivana, 2005 |
| corymbiferin-1- <i>O</i> -glucoside (19) | <i>G. amarella</i> ssp. <i>acuta</i> <i>G. bulgarica</i> <i>G. campestris</i> <i>G. crispatia</i> <i>G. ramosa</i> <i>G. amarella</i> ssp. <i>acuta</i> | Urbain et al, 2009 Janković, Krstić, and Ivana, 2005 Urbain et al, 2009 Janković, Krstić, and Ivana, 2005 Benn et al, 2009 Janković, Krstić, and Ivana, 2005 |
| corymbiferin-3- <i>O</i> -glucoside (20) | <i>G. campestris</i> <i>G. ramosa</i> <i>G. acuta</i> <i>G. albanica</i> <i>G. amarella</i> ssp. <i>acuta</i> | Urbain et al, 2009 Urbain et al, 2009 Urbain et al, 2009 Janković, Krstić, and Ivana, 2005 Urbain et al, 2009 |
| demethylbellidifolin (21) | <i>G. antarctica</i> <i>G. azurea</i> <i>G. bellidifolia</i> <i>G. bulgarica</i> <i>G. campestris</i> <i>G. cerina</i> <i>G. corymbifera</i> <i>G. crispatia</i> <i>G. florida</i> <i>G. germanica</i> (Willd.) Börner <i>G. multicaulis</i> <i>G. nitida</i> <i>G. nevadensis</i> <i>G. ramosa</i> <i>G. tenella</i> (Rottb.) Börner <i>G. parviflora</i> <i>G. patula</i> | Benn et al, 2009 Zhang and Yang, 1994 Carbonnier et al, 1977; Benn et al, 2009 Janković, Krstić, and Ivana, 2005 Carbonnier, Massias, and Molho, 1977; Urbain et al, 2009 Benn et al, 2009 Benn et al, 2009 Janković, Krstić, and Ivana, 2005 Nadinic et al, 1997 Carbonnier, Massias, and Molho, 1977 Hostettmann and Jacot, 1978 Rosella et al, 2007 Lacaille-Dubois and Galle, 1996 Báez, Quintero, and Nieves, 1999 Carbonnier, Massias, and Molho, 1977; Urbain et al, 2009 Carbonnier, Massias, and Molho, 1977 Rosella et al, 2007 Benn et al, 2009 |

To be continued

Continued Table 1

| Compounds | Species | References |
|--|--|---|
| demethylswertipunicoside (22) | <i>G. antarctica</i> <i>G. bellidifolia</i> <i>G. cerina</i> <i>G. corymbifera</i> <i>G. patula</i> <i>G. stenocalyx</i> Harry Sm. <i>G. australis</i> <i>G. antarctica</i> <i>G. corymbifera</i> <i>G. bellidifolia</i> <i>G. cerina</i> <i>G. corymbifera</i> <i>G. florida</i> <i>G. multicaulis</i> <i>G. patula</i> <i>G. parviflora</i> <i>G. serotina</i> <i>G. albanica</i> <i>G. bulgarica</i> <i>G. crispata</i> <i>G. acuta</i> <i>G. antarctica</i> <i>G. bellidifolia</i> <i>G. campestris</i> <i>G. caucasica</i> <i>G. cerina</i> <i>G. corymbifera</i> <i>G. germanica</i> <i>G. multicaulis</i> <i>G. nitida</i> <i>G. patula</i> <i>G. parviflora</i> <i>G. ramosa</i> <i>G. serotina</i> <i>G. stenocalyx</i> <i>G. thyrsoidae</i> <i>G. nitida</i> <i>G. nitida</i> <i>G. albanica</i> <i>G. amarellus</i> ssp. <i>acuta</i> <i>G. austriaca</i> <i>G. bellidifolia</i> <i>G. bulgarica</i> <i>G. campestris</i> <i>G. campestris</i> <i>G. crispata</i> <i>G. germanica</i> <i>G. multicaulis</i> <i>G. nitida</i> <i>G. parviflora</i> <i>G. ramosa</i> <i>G. thyrsoidae</i> <i>G. antarctica</i> <i>G. azurea</i> <i>G. bellidifolia</i> <i>G. cerina</i> <i>G. corymbifera</i> <i>G. florida</i> <i>G. multicaulis</i> <i>G. patula</i> <i>G. parviflora</i> <i>G. thyrsoidae</i> <i>G. amarellus</i> ssp. <i>acuta</i> <i>G. campestris</i> <i>G. ramosa</i> <i>G. amarellus</i> ssp. <i>acuta</i> <i>G. campestris</i> <i>G. ramosa</i> <i>G. nitida</i> <i>G. nitida</i> | Benn <i>et al.</i> , 2009 Benn <i>et al.</i> , 2009 Benn <i>et al.</i> , 2009 Benn <i>et al.</i> , 2009 Benn <i>et al.</i> , 2009 Massias, Carbone, and Molho, 1982 Nadinic, Penna, and Saavedra, 2002 Benn <i>et al.</i> , 2009 Benn <i>et al.</i> , 2009 Nadinic <i>et al.</i> , 1997 Rosella <i>et al.</i> , 2007 Benn <i>et al.</i> , 2009 Rosella <i>et al.</i> , 2007 Massias, Carbone, and Molho, 1982 Janković, Krstić, and Ivana, 2005 Janković, Krstić, and Ivana, 2005 Janković, Krstić, and Ivana, 2005 Lv and Li, 2009 Benn <i>et al.</i> , 2009 Benn <i>et al.</i> , 2009 Massias, Carbone, and Molho, 1982 Jensen and Schripsema, 2002 Benn <i>et al.</i> , 2009 Benn <i>et al.</i> , 2009 Hostettmann-Kalda and Jacot, 1978 Rosella <i>et al.</i> , 2007 Lacaille-Dubois and Galle, 1996 Benn <i>et al.</i> , 2009 Rosella <i>et al.</i> , 2007 Hostettmann-Kalda and Jacot, 1978 Massias, Carbone, and Molho, 1982 Massias, Carbone, and Molho, 1982 Tomas, Lock, and Jurupe, 1999 Lacaille-Dubois and Galle, 1996 Lacaille-Dubois and Galle, 1996 Janković, Krstić, and Ivana, 2005 Urbain <i>et al.</i> , 2009 Janković, Krstić, and Ivana, 2005 Benn <i>et al.</i> , 2009 Janković, Krstić, and Ivana, 2005 Benn <i>et al.</i> , 2009 Urbain <i>et al.</i> , 2009 Janković, Krstić, and Ivana, 2005 Benn <i>et al.</i> , 2009 Rosella <i>et al.</i> , 2007 Lacaille-Dubois and Galle, 1996 Rosella <i>et al.</i> , 2007 Urbain <i>et al.</i> , 2009 Tomas, Lock, and Jurupe, 1999 Benn <i>et al.</i> , 2009 Zhang and Yang, 1994 Benn <i>et al.</i> , 2009 Benn <i>et al.</i> , 2009 Benn <i>et al.</i> , 2009 Nadinic <i>et al.</i> , 1997 Rosella <i>et al.</i> , 2007 Benn <i>et al.</i> , 2009 Rosella <i>et al.</i> , 2007 Tomas, Lock, and Jurupe, 1999 Urbain <i>et al.</i> , 2009 Urbain <i>et al.</i> , 2009 Lacaille-Dubois and Galle, 1996 Lacaille-Dubois and Galle, 1996 |
| gentisein (23) | | |
| isobellidifolin (24) | | |
| lanceoside (25) | | |
| mangiferin (26) | | |
| norswertianine (27) | | |
| norswertianine 1- <i>O</i> -glucoside (28) | | |
| norswertianolin (29) | | |
| scherzerin (30) | | |
| swertiabissanthone-I (31) | | |
| swertiabissanthone-I 8'- <i>O</i> -glucoside (32) | | |
| swertianine (33) | | |
| swertianine 1- <i>O</i> -primeveroside (34) | | |

To be continued

Continued Table 1

| Compounds | Species | References |
|---|--------------------------------------|-----------------------------------|
| swertianine 8-O-glucoside (35) | <i>G. nitida</i> | Lacaille-Dubois and Galle, 1996 |
| swertianolin (36) | <i>G. acuta</i> | Lv and Li, 2009 |
| | <i>G. albanica</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. amarella</i> ssp. <i>acuta</i> | Urbain et al, 2009 |
| | <i>G. azurea</i> | Zhang and Yang, 1994 |
| | <i>G. bulgarica</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. campestris</i> | Urbain et al, 2009 |
| | <i>G. corymbifera</i> | Benn et al, 2009 |
| | <i>G. crispata</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. multicaulis</i> | Rosella et al, 2007 |
| | <i>G. parviflora</i> | Rosella et al, 2007 |
| | <i>G. ramosa</i> | Urbain et al, 2009 |
| swertipunicoside (37) | <i>G. antarctica</i> | Benn et al, 2009 |
| | <i>G. bellidifolia</i> | Benn et al, 2009 |
| | <i>G. cerina</i> | Benn et al, 2009 |
| | <i>G. corymbifera</i> | Benn et al, 2009 |
| | <i>G. patula</i> | Benn et al, 2009 |
| tripexanthoside C (38) | <i>G. amarella</i> ssp. <i>acuta</i> | Urbain et al, 2009 |
| | <i>G. campestris</i> | Urbain et al, 2009 |
| | <i>G. ramosa</i> | Urbain et al, 2009 |
| veratriloside (39) | <i>G. albanica</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. amarella</i> ssp. <i>acuta</i> | Urbain et al, 2009 |
| | <i>G. bulgarica</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. campestris</i> | Urbain et al, 2009 |
| | <i>G. crispata</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. ramosa</i> | Urbain et al, 2009 |

sesterterpenoids (nitidasin **45** and alborosin **46**) (Kawahara et al, 1997; Kawahara, Nozawa, and Flores, 2000), a flavonoid (luteolin **47**) (Lv and Li, 2009), a lignan glycoside (oivil 4'-O-glucose **48**), a sterol (daucosterol **49**), and a fatty alcohol (1-triacontanol **50**) (Zhang and Yang, 1994) were also isolated from the genus (Fig. 3).

Biological activities

Anti-oxidative and radioprotective properties

Some species of *Gentianella* Moench were reported to possess strong anti-oxidant properties, for example, *G. alborosea* and *G. nitida* extracts showed a noticeable radical scavenging activity by using DPPH assay (Rosario et al, 2004; Acero Llinas, and Galán, 2006). *G. austriaca* can significantly reduce lipid peroxidation (Leskovac et al, 2004), and the xanthones and flavone C-glucosides existed in *G. austriaca* may contribute to the anti-oxidative properties of the plant (Janković, Savikin, and Menkovic, 2008).

The species of *Gentianella* Moench also possessed radio-protective properties (Leskovac et al, 2004). In a recent study, the radioprotective effects of the different fractions of *G. austriaca* as well as the pure isolated compounds (demethylbellidifolin, demethylbellidifolin 8-O-glucoside, bellidifolin 8-O-glucoside, and swertisin) were investigated in irradiated human lymphocytes *in vitro*. The results suggested that the ethyl ether and

methanol fractions of *G. austriaca* showed significant protection in treatment of human lymphocytes after gamma-irradiation than the isolated compounds (Janković, Savikin, and Menkovic, 2008).

Antimicrobial activity

The crude ethanolic extract of *G. nitida* possessed mainly antifungal activity against *Candida albicans*, *Trichophyton mentagrophytes* and *Microsporum gypseum* (Rosario et al, 2004). Some studies reported that *G. achalensis* and *G. nevadensis* showed antibacterial activity against *Bacillus subtilis* (Báez, Quintero, and Nieves, 1999; Nadinic, Penna, and Saavedra, 2002). In addition, *G. alborosea* was reported to show strong inhibition of *Staphylococcus aureus* which might explain their efficacy in traditional acne treatment (Rainer, Bussmann, and Doris, 2008).

Anti-inflammatory activity

The dichloromethane extracts of *G. achalensis* showed marked anti-inflammatory activity in TPA-induced ear edema test (Nadinic et al, 1999).

Hypoglycemic activity

The crude extracts of *G. nitida* and *G. tristicha* (Gilg) Fabris ex T. N. Ho & S. W. Liu were reported to significantly lower the loaded glucose level in normal as well as alloxan-induced diabetic rats (Ninosca et al, 2001). The dichloromethane, methanol, and aqueous extracts of *G. thyrsoidea* also showed hypoglycemic and

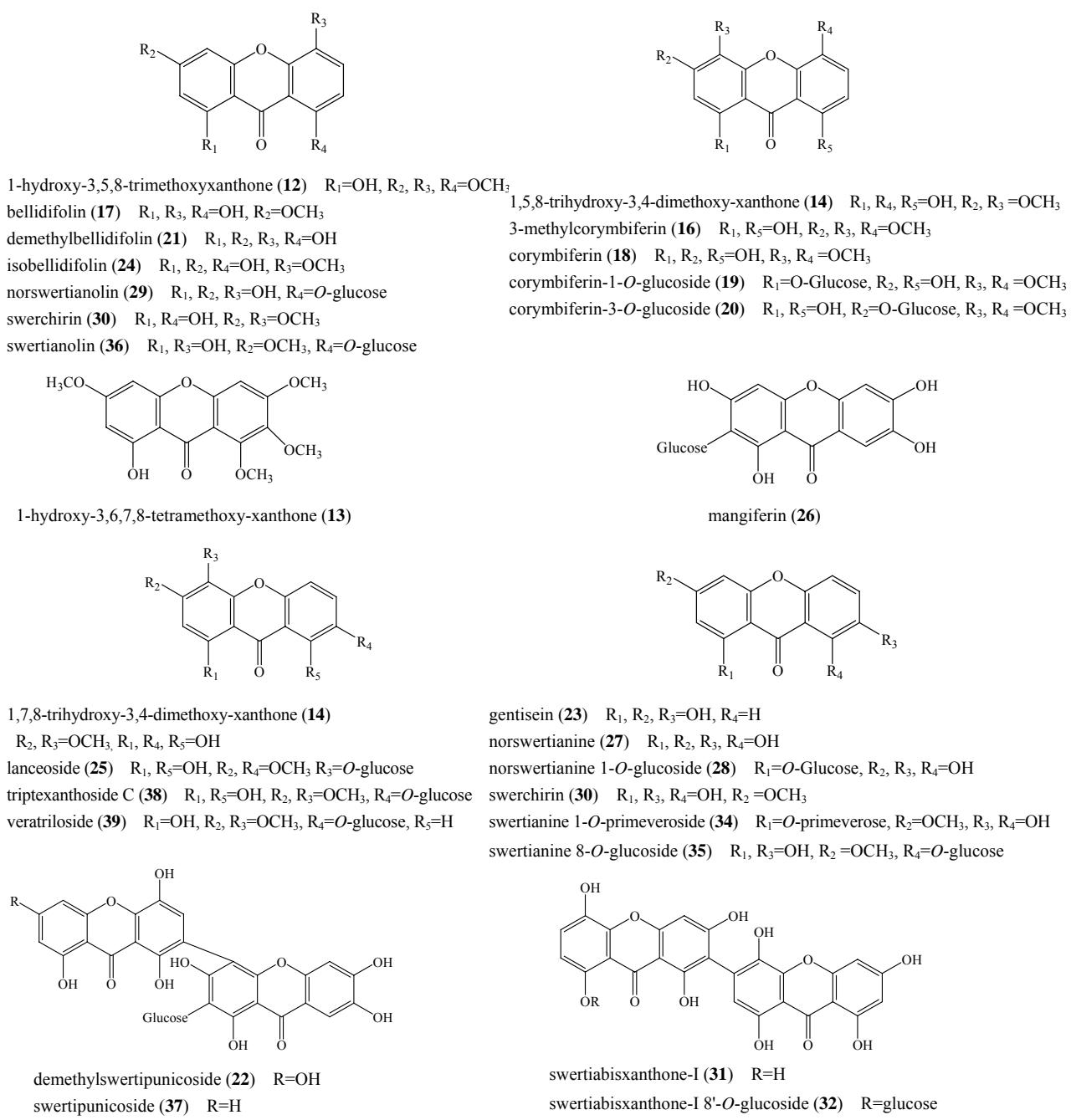


Fig. 2 Structures of xanthones in plants of *Gentianella Moench*

and hypocholesterolemic activities in rats (Tomas, Lock, and Jurupe, 1999). Bellidifolin, a common xanthone in plants of *Gentianella Moench*, was reported to be a potent hypoglycemic agent in STZ-induced diabetic rats by both oral and ip administration. This compound also lowered blood triglyceride levels significantly. It stimulated glucose uptake activity in Rat 1 fibroblasts expressing human insulin receptors (José *et al*, 1995). In Japan the extract of *G. alborosea* has been patented in the treatment of diabetes, obesity, and liver disorder, and

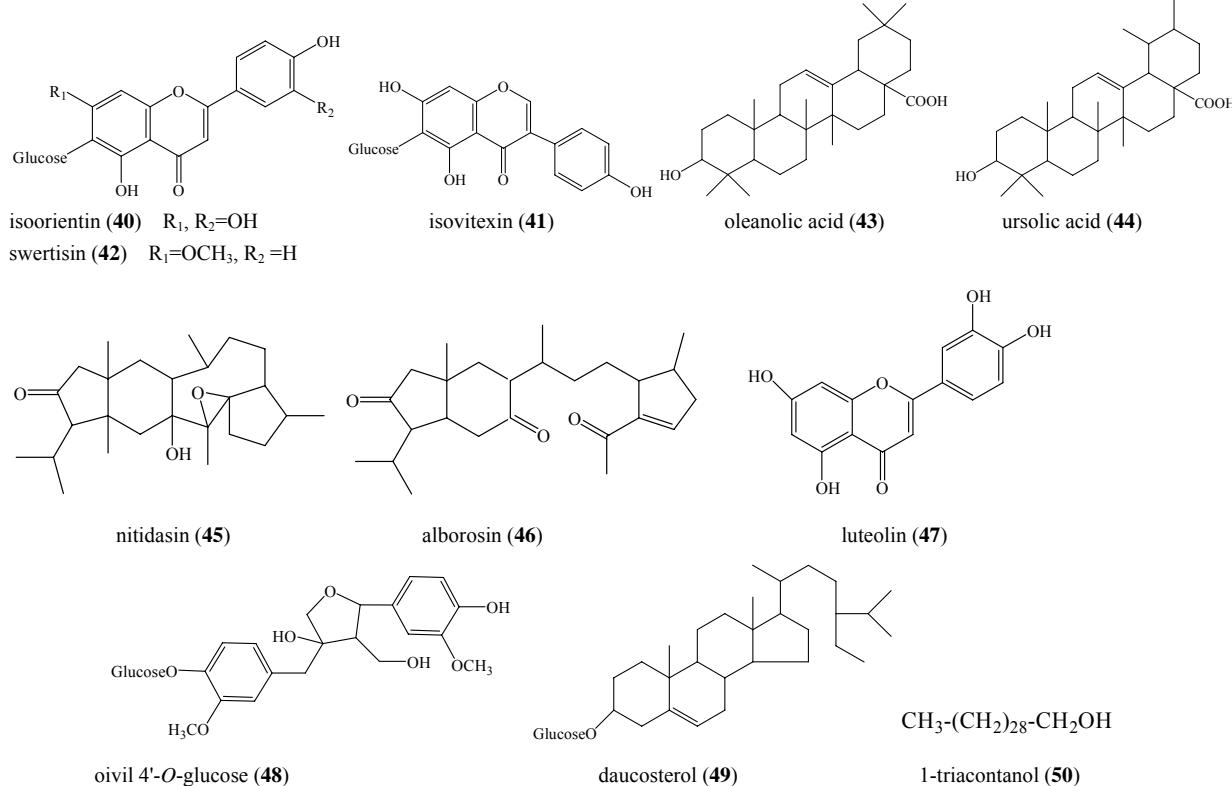
as health food for improving insulin resistance and decreasing blood sugar (Ogawa, 2006; 2007).

Anticholinergic activity

Swertiamarin and mangiferin, widely distributed in plants of *Gentianella Moench*, have shown anticholinergic properties (Bhattacharya *et al*, 1976; Yamahara *et al*, 1991). In a recent study, triptexanthoside C, isolated from *G. amarella* ssp. *acuta* also showed active against acetylcholinesterase, with an IC_{50} of $(13.8 \pm 1.6) \mu\text{mol/L}$ (Urbain *et al*, 2008).

Table 3 C-glucoflavonoids in plants of *Gentianella Moench*

| Compounds | Species | References |
|------------------|-----------------------|-----------------------------------|
| isoorientin (40) | <i>G. acuta</i> | Lv and Li, 2009 |
| | <i>G. albanica</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. azurea</i> | Zhang and Yang, 1994 |
| | <i>G. bulgarica</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. campestris</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. corymbifera</i> | Benn et al, 2009 |
| | <i>G. crispata</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. germanica</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. multicaulis</i> | Rosella et al, 2007 |
| | <i>G. nitida</i> | Lacaille-Dubois and Galle, 1996 |
| | <i>G. parviflora</i> | Rosella et al, 2007 |
| | <i>G. ramosa</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. serotina</i> | Massias, Carbone, and Molho, 1982 |
| isovitexin (41) | <i>G. corymbifera</i> | Benn et al, 2009 |
| | <i>G. multicaulis</i> | Rosella et al, 2007 |
| | <i>G. parviflora</i> | Rosella et al, 2007 |
| | <i>G. stenocalyx</i> | Massias, Carbone, and Molho, 1982 |
| swertisin (42) | <i>G. albanica</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. austriaca</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. bulgarica</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. crispata</i> | Janković, Krstić, and Ivana, 2005 |
| | <i>G. campestris</i> | Massias, Carbone, and Molho, 1982 |
| | <i>G. corymbifera</i> | Benn et al, 2009 |
| | <i>G. germanica</i> | Hostettmann and Jacot, 1978 |
| | <i>G. multicaulis</i> | Hostettmann and Jacot, 1978 |
| | <i>G. ramosa</i> | Rosella et al, 2007 |
| | <i>G. serotina</i> | Massias, Carbone, and Molho, 1982 |
| | <i>G. stenocalyx</i> | Massias, Carbone, and Molho, 1982 |
| | <i>G. parviflora</i> | Rosella et al, 2007 |

**Fig. 3** Structures of C-glucoflavonoids and other compounds in plants of *Gentianella Moench***Hepatoprotective activity**

Gentiopicroside, which is ubiquitous secoiridoid constituents in plants of *Gentianella Moench*, has shown hepatoprotective activities on the two hepatic injury models (carbon tetrachloride-induced and lipopolysaccharide-induced hepatitides) (Kondo et al, 1994), and is being used as an antihepatitis drug. Mangiferin and sweroside, two common constituents in plants of *Gentianella Moench* plants, were reported to show hepatoprotective activities (Zhou, 1991; Dar et al, 2005).

Other activities

Some studies reported bellidifolin and demethylbellidifolin isolated from species of *Gentianella Moench* were selective inhibitors of monoamine oxidases A enzymes (Báez, Quintero, and Nieves, 1999; Tovilović et al, 2005; Urbain et al, 2008), while swertianolin, the 8-O-lucopyranoside form of bellidifolin, gave 93.6% inhibition of MAO B activity at 10⁻⁵ mol/L (Urbain et al, 2008).

The extract of *G. alborosea* was reported to induce apoptosis on HeLa cells (ATCC and CCL-2), and the effects were dose-dependent (higher tested concentrations have the higher percentage of apoptotic nuclei) (Acero et al, 2006).

In addition, nitiol, a sesterterpenoid isolated from *G. nitida*, possessed enhancement activity of IL-2 gene expression in Jurkat cells (a human T cell line) (Kawahara *et al.*, 1999).

Toxicity

The toxicity of fractions obtained from extracts of *G. nevadensis* was tested using the brine shrimp (*Artemia salina*) assay. For ethyl acetate extract, the lowest values of median lethal concentrations were found in the fractions 4, 5, and 6 (72.97, 216.33, and 746.79 ppm, respectively), while for dichloromethane extract, fractions 2 and 3 showed higher toxicity against *A. salina* (LC_{50} 31.11 and 232.63 ppm, respectively) (Báez, Quintero, and Nieves, 1999).

It was reported that overdosing of *G. dianthoides* could cause miscarriage in pregnant women, and exceeding dosage of *G. graminea* can lead to blindness (Rainer and Bussmann, 2006).

Conclusion

In literature, only a few species among the 250 known from the plants of *Gentianella* Moench have been reported in their chemical investigations and pharmacological activities. But clearly, members of the plants of *Gentianella* Moench possess significant potentials for their pharmacological activities in the context of ethnomedicinal knowledge, especially in the treatments of heart diseases, diabetes, and hyperpoidemia. Eleven iridoids, 28 xanthones, three C-glucoflavonoids, and eight other compounds were isolated from the genus. Most of these compounds are associated with anti-crobial, anti-inflammatory, anti-oxidant, hypoglycemic, and antitumor activities, which provide an empirical base for the traditional utilization in the plants of *Gentinella* Moench. The extracts from many species are also added to many health-related products including diet pills, hair-loss products, homeopathic medicines, body cleansing products, and others. The species of *Gentianella* Moench described in this review do not appear to have sustainable supply issue and might serve as an important source of medicine in the world.

It is important to note that the species of *Gentianella* Moench species have not been fully explored concerning either safety or toxicity aspects. Only a little published data concerned the overdosing

and toxicity of some species. Therefore the toxicities of the traditional remedies and isolated chemical compounds should be further assessed.

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